# THE INFLUENCE OF NOVEL 4-THIAZOLIDINONE DERIVATIES IN CYTOPROTECTIVE MECHANISMS OF SMALL INTESTINE UNDER NSAID-INDUCED DAMAGE

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The aim of investigation was to compare the action of novel 4-thiazolidinone derivaties (compounds Les-5054 and Les-5055) toward parameters of nitroso-oxidative processes in mucous membrane of small intestine (MMSI) in rats on the background of indomethacin induced injury. The activity of nitric oxide synthases, myelopeoxidase, content of NO, and parameters of lipoperoxidation processes were measured in MMSI and the level of H2S and L-arginine in blood serum. Administration of indomethacin caused significant destructive damages in distal part of small intestine and increase in activity of inducible nitric oxide synthase (iNOS) and intensity of lipoperoxidation processes in comparison to control were observed. Also indomethacin injection was accompanied by decrease of  $H_2S$  and L-arginine level in blood serum. Administration of 4-thiazolidinone derivaties on the background of indomethacin induced injury reduce the activity of iNOS, myeloperoxidase, intensity of lipid peroxidation and increase generation of  $H_2S$ , that may be linked with the structure of this compounds. However compound Les-5054 showed more significant cytoprotective effect and antioxidant properties than compound Les-5055. Thus, the novel 4-thiazolidinone derivaties led to reduce of nitroso-oxidative processes caused by administration of NSAIDs.

*Key words: hydrogen sulfide, 4-thiazolidinones, small intestine, indomethacin-induced injury.* 

ydrogen sulfide (H<sub>2</sub>S) is now recognized as an important gasotransmitter together with nitric oxide (NO) and carbon monoxide (CO). H<sub>2</sub>S has been implicated in the induction of such processes as inhibition of leukocytes adherence to blood vessels, increase of endogeneous prostaglandins production in small intestine, induction of vasodilatation, increases cyclic AMP (cAMP) production in neural retina, modulates epithelial secretion and promotes resolution of colitis[1]. The deficiency of hydrogen sulfide could lead to various pathological changes in digestive tract, such as gastric mucosal injury, liver cirrhosis etc [2].

Nowadays nonsteroidal anti-inflammatory drugs (NSAIDs) are the most widely used class of drugs for treating inflammatory conditions such as: osteochondrosis, polyarthritis, headaches. However, they cause significant adverse reaction in the mucous membranes of the digestive system in the form of erosions, ulcers, impaired motor skills [3]. Owing to the inhibition of cyclooxygenase, NSAID suppress synthesis of prostaglandins which leads to leukocyte adherence to the vascular endothelium in the gastro-intestinal microcirculation, reduce bicarbonate and

mucus secretion by the epithelium, decrease blood flow, that play key role in the process of gastrointestinal injury caused by NSAIDs [4]. Also these drugs reduce  $H_2S$  generation by modulating the expression of activity of cystathionine- $\gamma$ -lyase, which is mainly expressed in smooth muscle cells in the cardiovascular system and in the gastrointestinal tract [5].

Thus, 4-thiazolidinones are one of important source of organic sulfurcontaining compounds and have been widely investigated regarding their therapeutic applications. Thiazolidinone-based molecules are attaractive targets in rational design of "druglike" compounds which possess anti-inflammatory, antioxidant, antitumor, choleretic, diuretic and other activities [6-8].

In the present study the effects of a novel 4-thiazolidinones (compounds Les-5054 and Les-5055), as potential H2S donors or mediators of its signaling pathways were investigated. The effects of these compounds in terms of nitroso-oxidative and cytoprotective effects, ability to decrease small intestinal injury, and acute anti-inflammatory effects were compared.

### **Materials and Methods**

Animals. The experimental protocols were approved by the Ethical Committee of Lviv National Medical University (Ukraine). Male albino rats weighing 200-250 g were used. The rats were fed standart chow and water ad libitum, and were housed in room with controlled temperature ( $22 \pm 1$  °C), humidity (65-70%) and light cycle (12 h light/dark).

The study comprised of the following series of experiments: 1 - intact animals were used as controls (n = 10); 2 - small intestine lesions in rats were induced by indomethacin (Sigma, USA) in dose of 35 mg/kg subcutaneously (n = 10) as previously described (n = 10) [9]; 3,4 - experimental groups, animals received compounds Les-5054, Les-5055 (10 mg/kg) per os once daily per 72 h on the background of indomethacin-induced injury (n = 10). The compounds Les-5054, Les-5055 were synthesized by prof. Lesyk R. in the Department of Pharmaceutical, Organic and Bioorganic Chemistry Danylo Halytsky Lviv National Medical University (Fig. 1) [8, 10].

Under general anesthesia, rats were sacrificed by decapitation and 10 sm of distal part of small intestine was then blindly evaluated for hemorrhagic damage. This involved measuring the lengths (mm) of all hemorrhagic lesions. The intestinal damage scores were then calculated by summing the lengths of all lesions for each rats. The mucous membrane of small intestine (MMSI) samples were homogenized in phosphate buffer pH 6.0 1:4 and centrifuged at 3000 rpm, supernatant was used to determine values of biochemical parameters.

Determination of NO-system in mucous membrane of small intestine. The content of NO in homogenate was determined as nitrites by the method of Green et al. [11]. The absorbance was read in a Stat fax at 550 nm. NO concentration was expressed as mmol/g. NO-synthases (general NOS, iNOS, and eNOS) activity was measured by the method described in detail [12]. NOS activity was expressed in nmol L-citrylline/min×mg of protein.

Measurement of L-arginine and H<sub>2</sub>S in blood serum. The level of L-arginine in plasma samples was measured by Sakaguchi reaction [13]. Plasma L-arginine level was expressed as mmol/l. H<sub>2</sub>S concentration was determined by reaction with N,N-dimethyl-para-phenylenediamine in the presence of FeCl<sub>2</sub> and expressed as mmol/g [14].

Lipid peroxidation determination. Lipid peroxidation level was expressed as MDA (malonic dial-

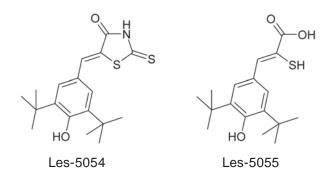


Fig. 1. Structures of 4-thiazolidinone derivatives

dehyde) concentration in homogenates of MMSI. It was measured according to the procedure of Timirbulatow et al. [15]. MDA levels were expressed as mmol/l.

Intracellular myeloperoxidase activity. Myeloperoxidase (MPO) content in homogenates of MMSI measured at 460 nm according to the procedure of Bradley et al. [16]. MPO level were expressed as U/mg.

Antioxidant enzymes defence determination. Activity of superoxide dismutase (SOD) was determined by the reaction of reduction of nitrotetrazoliume blue to nitroformazan [17]. SOD activity was expressed in mmol/min×mg of protein. Catalase (CAT) activity was determined by measuring of the decrease in hydrogen peroxide concentration at 410 nm by the Korolyuk method [18]. Colon mucosal catalase activity was expressed in mmol  $\rm H_2O_2/min\times mg$  of protein.

Statistics. The statistical processing of the data was done by conventional methods for analysis of variance using MS Excel software for Student's t-test. The difference was considered to be significant at P < 0.05.

## **Results and Discussion**

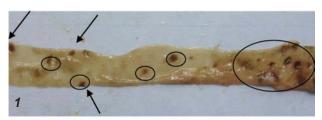
In our study, injection of indomethacin (35 mg/kg) manifested by erosions and hemorrhages, with a total area of  $74 \pm 5,94$  mm<sup>2</sup> (Fig. 2, 3). Indometacin induced injury in the MMSI was associated with change of the activity of NO-synthases: activity of general NOS decreased (from  $815.5 \pm 49.8$  to  $595.54 \pm 73.7$  nmol/min×mg) (P < 0.05), activity of eNOS was decreased by 55% (P < 0.01), and activity of iNOS increased more than threefold (P < 0.01) as compared with indeces of control group. In MMSI concentration of NO was markedly elevated in two times and, concomitantly (P < 0.01), content of L-ar-

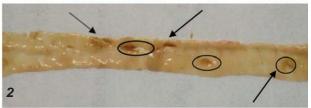
ginine in serum blood decreased for 33% (P < 0.05). It was found that H<sub>2</sub>S formation from L-cysteine was dependently inhibited by addition of indomethacin by 10% (P < 0.05) (Table 1).

MMSI, affected with indometacin induced injury, was subjected to the following changes: enhanced activity of lipoperoxidation processes manifested by a steep rise of MDA concentration – for 56% (P < 0.01) at that, MPO activity enhanced more than 4-fold (P < 0.01), and catalase activity – for 32% (P < 0.01). The activity of SOD was not statistically significant (Table 2).

In the presence study the development of NSAID-induced small intestinal damage via indomethacin injection was acompanied by enhanced processes of lipid peroxidation, increase activity of iNOS and myeloperoxidase that led to development of hemorrhagic lesions in distal part of small intestine. This destructive changes caused by inflammation processes and acomplished throught decrease contents of  $H_2S$  which enhance gastrointestinal mucosal resistance to injury [19].

On another hand, the injection of NSAIDs such as diclofenac, naproxen, indomethacin caused the non selective inhibition of both COX-1 and COX-2 and, as result, a significant reduction in production of prostaglandins that is the key factor in development of enteropathies.





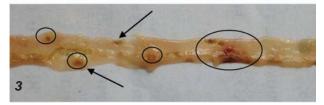


Fig. 2. Representative photographs of the small intestinal injury: 1 – indomethacin; 2 – Les-5054 + indomethacin; 3 – Les-5055 + indomethacin

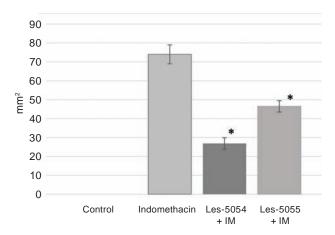


Fig. 3. The area of the structure-hemorrhagic damage of the MMSI. \*P < 0.05 versus the indexes in indomethacin (IM)

Compound Les-5054 displayed significant cytoprotective effect that manifested by separate hemorrhages with the absence of considerable destructive changes of the MMSI. The total area of hemorrhagic lessins decreased for 63% (P < 0.05) (Fig. 2, 3). The administration of Les-5054 on the background of indomethacin-induced injury decrease the activity of iNOS for 35% (P < 0.01), and activity of eNOS increased for 52% (P < 0.01) as compared with independent action of indomethacin. Intensity of lipoperoxidation processes were determined much lower than under the effect of indomethacine, MDA concentration declined for 32% (P < 0.01). Administration of compound Les-5054 reversed the inhibition of H<sub>2</sub>S caused by indomethacin and increase it for 24% (P < 0.05) as compared with indices of the second group.

Compound Les-5054 led to 3 time decreased MPO activity as compared with indices of 2 group (P < 0.05). It was found the administration of compounds Les-5054 and Les-5055 as  $H_2S$  releasing compounds can inhibit peroxidase activity of MPO, key role is production of reactive oxidants (HOCl etc) which influence to a healthy tissue is the main indication of MPO's pathophysiological functions [20].

In our investigations it was shown that administration of Les-5055 on the background of indomethacin-induced injury also shows cytoprotective effect but not as much as compound Les-5054. The total area of hemorrhagic lessins decreased for 37% (P < 0.05) as compared with independent action of indomethacin (Fig 2, 3). Activity of eNO-synthases had a tendency to increase, whereas iNOS activity

Table 1. Effect of novel 4-thiazolidinones at the background of indomethacin-induced injury on concentration of malonic dialdehyde, nitrite anion, activity of nitric oxide synthases and arginase in MMSI and concentration of  $H_2S$  and L-arginine in serum blood

Variable	Experimental Groups				
	Control group	Indomethacin 35 mg/kg	Les-5054 + Indomethacin	Les-5055 + Indomethacin	
Malonic dialdehyde,					
(µmol/g)	$186.6 \pm 8.1$	$291.4 \pm 26.7**$	$199.1 \pm 11.1^{##}$	$242.6 \pm 16.0$ <sup>##</sup>	
Nitrite anion, (µmol/g)	$1.2 \pm 0.1$	$2.8\pm0.2 \textcolor{red}{**}$	$1.8 \pm 0.2$	$1.9 \pm 0.1$	
Total nitric oxide synthase – NOS, (nmol/min·g) Inducible nitric oxide	$815.5 \pm 49.8$	$595.5 \pm 73.1$	$637.9 \pm 42.8$	$578.2 \pm 64.7$	
synthase – iNOS, (nmol/min·g) Constitutive nitric	$66.1 \pm 24.9$	203.6 ± 26.8**	132.9 ± 27.5##	162.3 ± 27.7#	
oxide synthase – cNOS,	<b>50</b> 0 6 + 664	224.0 (2. 5/14)	<b>5</b> 0.60 . <b>15</b> 0##	44.70 . 44.04	
(nmol/min·g)	$728.6 \pm 66.1$	$331.9 \pm 62.5**$	$506.3 \pm 45.3^{##}$	$415.9 \pm 44.0^{\#}$	
Arginase, (µmol/ min·g)	$0.20 \pm 0.03$	$0.05 \pm 0.02**$	$0.15 \pm 0.04$ #	$0.09 \pm 0.006$	
L-Arginine, (µg/ml)	$46.7\pm3.6$	$31.2\pm2.8$	$42.6 \pm 4.1^{\#}$	$40.7 \pm 4.2$	
H <sub>2</sub> S (μmol/g×min)	$88.4 \pm 2.7$	$79.5 \pm 1.0*$	$98.5\pm2.6^{\#}$	$93.0\pm2.5^{\#}$	

Here and for table 2 results are expressed as mean  $\pm$  SD for 10 rats per group; \*P < 0.05, \*\*P < 0.01 in comparison of control group; \*P < 0.05, \*\*P < 0.01 versus the indices of indomethacin action.

Table 2. The activity of myeloperoxidase and antioxidant enzymes in MMSI of rats injected indomethacin with investigated compounds

Variable	Experimental Groups				
	Control group	Indomethacin	Les-5054 +	Les-5055 +	
		35 mg/kg	Indomethacin	Indomethacin	
SOD, (mmol/min×mg)	$23.9\pm1.0$	$27.8 \pm 1.3$	$24.9\pm0.9\#$	$27.9 \pm 1.5$	
CAT, (mmol H <sub>2</sub> O <sub>2</sub> /min×mg)	$16.9\pm1.6$	$22.4 \pm 2.6 \textcolor{red}{**}$	$20.9\pm1.2$	$20.0\pm2.4$	
MPO, (U/mg)	$1.2\pm0.3$	$5.0\pm0.5*$	$1.6\pm0.5\#$	$4.3 \pm 1.2$	

was reduced for 20% (P < 0.05), compared to their activity in indomethacin induced damages. Contents of NO and MDA also showed a tendency to decrease (P < 0.05).

Many reports showed that several H<sub>2</sub>S-based therapeutics corresponded to target disorders and were characterized by oxidative stress and associated tissue injury. It was shown that their cytoprotection was accompanied by the decrease of mRNA expression for pro inflammatory cytokines, such as IL-10 or TGF-b. They also prevent inflammation by decrease of MDA content, increase GSH level and

decrease of NO, IL-6 and TNF-a secretion in intestine mucosa [1].

In experimental model of indomethacin-induced injury characteristic damages in distal part of small intestine was observed. In our investigation we demonstrated that novel 4-thiazolidinones displayed significant cytoprotective effect, manifested by the decreased area of the MMSI lesions. Normalization of NO-synthases activities and the intensity of lipoperoxidation processes were found. Also administration of investigated compounds was associated with increase of H<sub>2</sub>S level in serum blood. Thus,

novel 4-thiazolidinones showed cytoprotective, antiinflammatory effects and antioxidant properties, and may be promising substances for new pharmacological preparations.

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#### References

- Magierowski M, Magierowska K, Kwiecien S, Brzozowski T. Gaseous Mediators Nitric Oxide and Hydrogen Sulfide in the Mechanism of Gastrointestinal Integrity, Protection and Ulcer Healing. *Molecules*. 2015; 20(5): 9099-9123.
- 2. Geng B, Chang L, Pan C, Qi Y, Zhao J, Pang Y, Du J, Tang C. Endogenous hydrogen sulfide regulation of myocardial injury induced by isoproterenol. *Biochem Biophys Res Commun*. 2004; 318(3): 756-763.
- 3. Fomenko I, Sklyarov A, Bondarchuk T, Biletska L, Panasyuk N, Wallace JL. Effects of conventional and hydrogen sulfide-releasing non-steroidal anti-inflammatory drugs in rats with stress-induced and epinephrine-induced gastric damage. *Stress.* 2014; 17(6): 528-537.
- 4. Gu X, Zhu YZ. Therapeutic applications of organosulfur compounds as novel hydrogen sulfide donors and/or mediators. *Expert Rev Clin Pharmacol.* 2011; 4(1): 123-133.
- 5. Fiorucci S, Antonelli E, Distrutti E, Rizzo G, Mencarelli A, Orlandi S, Zanardo R, Renga B, Sante M, Morelli A, Cirino G, Wallace JL. Inhibition of hydrogen sulfide generation contributes to gastric injury caused by anti-inflammatory nonsteroidal drugs. *Gastroenterol*. 2005; 129(4): 1210-1224.
- Geronikaki AA, Lagunin AA, Hadjipavlou-Litina DI, Eleftheriou PT, Filimonov DA, Poroikov VV, Alam I, Saxena AK. Computeraided discovery of anti-inflammatory thiazolidinones with dual cyclooxygenase/ lipoxygenase inhibition. *J Med Chem.* 2008; 51(6): 1601-1609.

- 7. Lozynskyi AV, Kaminskyy DV, Romanchyshyn KB, Semenciv NG, Ogurtsov VV, Nektegayev IO, Lesyk RB. Screening of antioxidant and anti-inflammatory activities among thiopyrano[2,3-d]thiazoles. *Biopolym Cell*. 2015; 31(2): 131-137.
- Lesyk RB, Zimenkovsky BS. 4-Thiazolidones: centenarian history, current status and perspectives for modern organic and medicinal chemistry. *Curr Org Chem.* 2004; 8(16): 1547-1577.
- 9. Yarushkina NI, Bagaeva TR, Filaretova LP. Somatic pain sensitivity under indometacin induced gastric and small intestinal injury in rats. *Rus Phisiol Zhurn*. 2014; 100(1): 73-85. (In Russian).
- 10. Kumar KSS, Hanumappa A, Vetrivel M, Hegde M, Girish YR, Byregowda TR, Rao S, Raghavan SC, Rangappa KS. Antiproliferative and tumor inhibitory studies of 2, 3 disubstituted 4-thiazolidinone derivatives. *Bioorg Med Chem Lett.* 2015; 25(17): 3616-3620.
- 12. Sumbajev VV, Yasinskaya IM. The influence of DDT on nitric oxide synthase activity in liver, lungs and brain of rats. *Modern Probl Toxycol*. 2000; 3: 3-7. (In Russian).
- 13. Alejnikova TL, Rubtsova GV, Pavlova NA. Manuals for practical lessons in biochemistry. M.: Medicine, 2000. 128 p. (In Russian).
- 14. Olkhovskiy OS, Zaichko N.. Influence propargyl glycine and sodium content of hydrogen and H<sub>2</sub>S-indices of antioxidant system in the myocardium of rats of different ages. Med Chem. 2013; 15(4): 10-15. (In Ukrainian).
- 15. Timirbulatov MA, Seleznev EI. Method for increasing the intensity of free radical oxidation of lipid-containing components of the blood and its diagnostic significance. *Lab Delo.* 1981; 4: 209-211. (In Russian).
- 16. Bradley PP, Priebat DA, Christensen RD, Rothstein G. Measurement of cutaneous inflammation: estimation of neutrophil content with an enzyme marker. *J Invest Dermatol*. 1982; 78(3): 206-209.
- 17. Chevari S, Andyal T, Shtrenger Ya. Determination of blood parameters and their role for diagnostics in elderly age. *Lab Delo.* 1991; 10: 9-13. (In Russian).

- 18. Koroluk M, Ivanova L, Mayorova I, Tokorev W. Method of determination of catalase activity. *Lab Techniq.* 1988; 1: 16-19. (In Russian).
- 19. Sklyarov AY, Lesyk RB, Panasyuk NB, Fomenko IS, Havrylyuk DY. Comparison of dual acting drugs and conventional NSAIDs towards parameters of NO-synthase system and oxidative stress in mucosal membrane of large
- intestine of rats with experimental ulcerative colitis. *Biopolym Cell*. 2011; 27(2): 147-153.
- 20. Pálinkás Z, Furtmüller PG, Nagy A, Jakopitsch C, Pirker KF, Magierowski M, Jasnos K, Wallace JL, Obinger C, Nagy P. Interactions of hydrogen sulfide with myeloperoxidase. *Br J Pharmacol.* 2015; 172(6): 1516-1532.

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