THE INDICES OF NITROGEN (II) OXIDE SYSTEM IN EXPERIMENTAL HEPATOPULMONARY SYNDROME

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Hepatopulmonary syndrome (HPS) is a pulmonary complication of liver disease characterized by arterial hypoxemia. Altered nitrogen (II) oxide (NO) production has also been implicated in the pathogenesis of the HPS. The present study was designed to evaluate the indices of NO system in the blood serum and lung tissue of animals with different models of hepatopulmonary syndrome. The total NOS activity was performed by monitoring the rate of conversion of L-arginine into citrulline. The total contents of NO metabolites was assessed by evaluation of their amount, which included nitrite ions that were previously presented in the sample (NO2–) and also nitrate ions reducted to nitrites (NO3–). We found a significant increase in total NOS activity in the lung tissue of rats of both experimental groups as compared to control animals, but it was greater in the rats on the 28th day after the common bile duct ligation. The total concentration of NO2– + NO3– in the lung tissue of the rats in the experimental group N 1 also significantly increased (5.8 times) and in the rats of the experimental group with carbon – 4.5 times (P < 0.001) vs the control group. Thus, in rats with different models of hepatopulmonary syndrome the activation of nitroxydergic process by a significant increase in nitrogen (II) oxide metabolites contents and total NO synthases activity has been established. Herewith a more pronounced intensification of nitroxydergic processes was observed in rats on the 28th day after the common bile duct ligation.

Key words: nitrogen (II) oxide, NO-synthase, rats, hepatopulmonary syndrome.

Liver cirrhosis has emerged as a major cause of global health burden. According to the Global Burden of Disease 2010 study, liver cirrhosis caused 31 million Disability Adjusted Life Years (DALYs), or 1.2% of global DALYs, in 2010, and one million deaths, or 2% of all deaths worldwide in that year [1]. In Ukraine the proportion of the deaths due to the digestive system disorders, which now occupy the fourth position in the structure of death causes, including 71.0% of patients with liver fibrosis and liver cirrhosis has also increased in recent years [2].

Hepatopulmonary syndrome (HPS) is a pulmonary complication of chronic liver disease characterized by arterial hypoxemia. This condition often manifests in adult patients with terminal liver disease, having a prevalence of 4 to 32% [3]. By other data, the prevalence of HPS varies from 4 to 47%, depending on the population and the criteria used to diagnose [4]. It is distinguished by three specific clinical entities consisting of liver disease and/or portal hypertension, disturbance of alveolar-arterial oxygen gradient (> 15 mm Hg or > 20 mmHg when age ≥ 65 years), and intrapulmonary vascular dilatations (IPVD) [4, 5].

Patients with progressive HPS have respiratory symptoms including, shortness of breath, clubbed fingers, and cyanosis. Although it commonly occurs, shortness of breath is a nonspecific symptom of HPS. The most common manifestations of HPS are platypnea (increased shortness of breath when the body is in a vertical position) and orthodeoxia (3-10 mmHg reduction in PaO2 in capillary blood during transition from horizontal to vertical position) [6].

The pathogenesis of HPS is not completely understood, and no effective pharmacological treatment has been developed yet [7]. Without a liver trans-
plant the prognosis of HPS is poor. If HPS develops, the risk of death in the next year is about 41% [8].

Since the basis of HPS pathogenesis is the dilatation of inner lung capillaries, researchers suggest that HPS is caused by the prolonged action of biologically active compounds on the blood vessels of pulmonary circuit. A possible role in resistant vasodilation have been suggested for many substances synthesized in the body. Potential mediators of HPS include: nitrogen (II) oxide, endothelin B and endothelin–I, prostaglandins E1 and I2, tumor necrosis factor–α, vasoactive intestinal polypeptide, substance P, calcitonin, glucagon, platelets activating factor, carbon monoxide and others [9].

The present study was, therefore, designed to evaluate the indices of nitrogen (II) oxide system in blood serum and lung tissue supernatant of animals with different models of hepatopulmonary syndrome.

Materials and Methods

The experiments were performed on 56 white nonlinear mature male rats, 180-220 g in weight that were housed at 25 ± 3 °C and humidity of 55 ± 2%, under a constant 12 h light and dark cycle. Water was available ad libitum.

All experiments were conducted in accordance with the European Convention for the protection of vertebrate animals used for experimental and other scientific purposes [10].

The experimental animals were divided into the 4 groups: I – control group N 1 (n = 12); II – experimental group N 1 (n = 18); III – control group N 2 (n = 12); IV – experimental group N 2 (n = 14).

The first experimental model of hepatopulmonary syndrome (HPS) was made by imposition of double ligature on common bile duct and its further dissection with a scalpel [11]. In the control group of animals N 1, the common bile duct was separated from the tissue, but not dissected. Postoperative wound was sewed up completely in layers. On the 28th day after the surgery the animals were taken out of the experiment under thiopental anaesthesia.

The animals of the second experimental group were fed a mixture of maize flour, lard, cholesterol, and alcohol plus subcutaneous injection with carbon tetrachloride (CCl4) oil solution for 8 weeks. The CCl4 oil solution (400 g/l) was injected at 0.5 ml/100 g body weight on the first day of the experiment and at 0.3 ml/100 g body weight from the third day at an interval of two days until the experiment end. Lard was used only in the first two weeks accounting for 20% of the feed. Cholesterol was appended at 0.5% of feed for the whole experiment. Alcohol was used in the drinking water exclusively (300 ml/l) during the whole experiment [12].

The control group of animals N 2 was on a standard diet of the vivarium and was administered intragastrically the equivalent amount of olive oil. The main advantage of this model is its non-invasiveness and multifactoriality, which makes it closer to the real causes in patients.

During the simulation of the HPS 8 animals died.

NOS activity assay in the lung tissue supernatant was performed by monitoring the rate of conversion of L-arginine into citrulline [13]. Total protein was measured by Lowry assay [14].

Briefly, the samples aliquots that contained 300 mg of protein were used to determine the total NOS activity. They were incubated for 60 min at 37 °C in a total volume of 1 ml substrate mixture (pH 7.0) of the following composition (µmol/ml): KH2PO4 – 50, MgCl2 – 1, CaCl2 – 2, NADPH (Sigma, USA) – 1, L-arginine – 2. The reaction was stopped by adding 0.3 ml of HClO4 (CNa = 2 mol/l). The samples that contained the full substrate mixture previously denatured by HClO4 (CNa = 2 mol/l) were used as a control. The mixture was centrifuged at 3500 g for 10 min and the non-protein supernatant mixtures were used to test L-citrulline by highly specific method for color reaction with antipyrine. Its sensitivity is 0.2 mg of L-citrulline in 1 ml, so it can be used to study the NOS activity. Protein-free aliquot samples were mixed with 2 ml of reagent (1 ml of 59 mmol/l diacetyl monoxime (Sigma, USA) + 1 ml of 32 mmol/l antipyrine (Sigma, USA) + 55 µmol/l Ferrous (II) sulphate in H2SO4 (CNa = 6 mol/l)) and boiled for 15 min in a water bath. After cooling the value of extinction was determined at 456 nm. The citrulline content was determined using a calibration graph. Total NOS activity was expressed as pmol of L-citrulline/min per 1 mg of protein.

Quantitative assessment of total concentration of NO2– + NO3– was performed by evaluation of their amount, which included nitrite ions that were previously presented in the sample (NO2–) as well as nitrate ions reduced to nitrites (NO3–) [15]. The reduction was performed using zinc dust in acidic environment. Nitrites with sulphanilic acid underwent a reaction of diazotisation. The obtained diazo-
isation compound with N-1-naphthylethylenediamin formed the azo dye. Optical density of the obtained colour solution was evaluated by spectrophotometry at 536 nm.

The total concentration of $\text{NO}_2^- + \text{NO}_3^-$ in the studied sample was estimated by the equation: $X = (Y - A)/B$, where $X$ – concentration of NO metabolites in μmol/l; $Y$ – optical density of the studied sample; $B$ – regression coefficient; $A$ – intercept.

All the data were processed using the software package Statistica 6.1 for Windows. Intergroup comparisons were performed using Mann–Whitney–Wilcoxon U test. The median (Me) and interquartile range (IQR [Q25–Q75]) were deduced. Differences with $P$-value < 0.05 were considered as significant.

**Results and Discussion**

Total NOS enzymatic activity in the lung tissue supernatant of rats with HPS was significantly increased in both experimental groups as compared to control animals, but was greater in the rats of the first experimental group (on the 28th day after the common bile duct ligation) (Table).

The total concentration of $\text{NO}_2^- + \text{NO}_3^-$ in blood serum of the rats on the 28th day after the common bile duct ligation was significantly increased (3.9 times, $P < 0.001$) vs control group N 1. In the rats of the second experimental group (with carbon tetrachloride induced cirrhosis) the total concentration of $\text{NO}_2^- + \text{NO}_3^-$ in blood serum also significantly increased (3.1 times, $P < 0.001$) vs control group N 2.

A comparison of total concentration of $\text{NO}_2^- + \text{NO}_3^-$ in blood serum and lung tissue supernatant was of great importance. It was determined that NO production disorders took place unidirectionally towards the oxidative burst. Thus, the total concentration of $\text{NO}_2^- + \text{NO}_3^-$ in lung tissue supernatant in the rats of the experimental group N 1 also increased 5.8 times ($P < 0.001$) and in the rats of the experimental group N 2 – 4.5 times ($P < 0.001$). When compared the total concentration of $\text{NO}_2^- + \text{NO}_3^-$ in 2 experimental models of HPS, its prevalence in rats on the 28th day after the common bile duct ligation was determined.

Since nitrogen (II) oxide (NO) is recognized as one of the most powerful endogenous pulmonary vasodilators, it has been suggested as the most likely candidate not only for the hyperdynamic type of circulation in liver cirrhosis, but also for HPS [16].

### The indices of nitrogen (II) oxide system in blood serum and lung tissue supernatant of rats with hepatopulmonary syndrome, Me [Q25–Q75]

<table>
<thead>
<tr>
<th>Group of animals</th>
<th>Control group N 1 ($n = 12$)</th>
<th>Experimental group N 1 ($n = 12$)</th>
<th>Control group N 2 ($n = 12$)</th>
<th>Experimental group N 2 ($n = 12$)</th>
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</thead>
<tbody>
<tr>
<td><strong>Blood serum</strong></td>
<td></td>
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<tr>
<td>$\text{NO}_2^- + \text{NO}_3^-$, μmol/l</td>
<td>36.7 [32.8; 38.7]</td>
<td>143.4 [131.6; 152.8]</td>
<td>33.4 [29.7; 35.9]</td>
<td>104.2 [96.3; 109.1]</td>
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<tr>
<td></td>
<td>$P_1 &lt; 0.001$</td>
<td>$P_1 &lt; 0.001$</td>
<td>$P_2 &lt; 0.05$</td>
<td>$P_1 &lt; 0.001; P_2 &lt; 0.05$</td>
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<tr>
<td><strong>Supernatant of lung tissue</strong></td>
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<td></td>
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<tr>
<td>$\text{NO}_2^- + \text{NO}_3^-$, μmol/l</td>
<td>14.1 [11.5; 16.4]</td>
<td>81.7 [77.5; 83.4]</td>
<td>12.0 [10.8; 13.4]</td>
<td>54.7 [50.8; 57.3]</td>
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<td></td>
<td>$P_1 &lt; 0.001$</td>
<td>$P_1 &lt; 0.001$</td>
<td>$P_1 &lt; 0.001; P_2 &lt; 0.05$</td>
<td>$P_1 &lt; 0.001; P_2 &lt; 0.05$</td>
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<tr>
<td>Total NOS, pmol of L-citrulline/min per mg of protein</td>
<td>46 [39; 52]</td>
<td>139 [117; 161]</td>
<td>51 [43; 58]</td>
<td>116 [101; 125]</td>
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<tr>
<td></td>
<td>$P_1 &lt; 0.001$</td>
<td>$P_1 &lt; 0.001; P_2 &lt; 0.05$</td>
<td>$P_1 &lt; 0.001; P_2 &lt; 0.05$</td>
<td>$P_1 &lt; 0.001; P_2 &lt; 0.05$</td>
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Notes: $P_1$ – significant difference as compared to the control animals; $P_2$ – significant difference between experimental animals.
tion in hepatocytes and Kupffer cells cause migration of intestinal organisms and endotoxins into the blood, causing bacteremia and intestinal endotoxemia. Endotoxins destroy mitochondria and lysosomes of intestinal epithelial cells, triggering cell autolysis. Ultimately, a vicious circle of intestinal endotoxemia and increased permeability of intestinal mucosa is created [9].

Zhang et al. showed a significant increase of endotoxins and TNF–α level in plasma and the increased number of Gram–negative microorganism colonies in rats with HPS that suggests that intestinal endotoxemia is indeed implicated in the molecular mechanisms of experimental HPS [17].

As a result of the appearance of portosystemic shunts and a decrease in the hepatic phagocytic capacity the lung filters the systemic blood to compensate the decrease in hepatic phagocytosis, and the increase in the lung phagocytic activity results in macrophage accumulation in the pulmonary endothelium [18].

So endotoxin induced activation of monocyte/macrophagal system involves Kupffer cells, macrophages of the spleen, pulmonary intravascular macrophages and blood mononuclear cells [19], and inducible NO synthase is a key enzyme in the macrophage that is potently induced in response to proinflammatory stimuli.

Our findings of increased nitric oxide production in case of HPS coincide with the studies of other authors. A series of experimental studies investigated NO quantities in the context of liver cirrhosis and HPS. Fallon et al. have emphasised the role of NO in experimental models of liver cirrhosis, where overexpression of eNOS by pulmonary vessels causes the increased production of endothelin–1 (ET–1) by cholangiocytes, resulting in the increased expression of endothelin receptor type B to ET–1 at the level of pulmonary vessels, and increases synthesis of nitrogen (II) oxide [20].

The level of NO in exhaled air was elevated in patients with HPS, returning to normal 3 to 12 months after the liver transplantation [21, 22]. In a similar study, B. Degano et al. found that concentration of NO in exhaled air of patients with cirrhosis was three times higher than that in healthy persons [23].

Using flow cytometry, which allowed differentiating alveolar and bronchial origin of NO, it was determined that the increased formation of NO mainly took place in the alveoli [24]. Notably, the production of nitrogen (II) oxide by alveoli can play an important role in haemodynamic disturbances and changes in gas exchange in patients with cirrhosis. Thus, a direct correlation between the alveolar production of NO and hyperdynamic type of circulation was established [24]. Moreover, overexpression of both inducible and constitutional isoforms of NO-synthase in alveolar macrophages and pulmonary endothelial cells was observed in rats with experimental liver cirrhosis [25, 26].

While this pulmonary overexpression has not yet been found in patients with HPS, it probably occurs in most cases of HPS, at least in patients who have been successfully treated using inhibitors of nitrogen (II) oxide synthesis or its targets, the so-called secondary messengers of cyclic guanosine monophosphate. Several studies successfully used the following therapeutic approaches in patients with HPS: either intravenous introduction of methylene blue, which is the main inhibitor of NO molecular target, cyclic guanosine monophosphate or spraying with NO synthase inhibitor (N-nitro-L-arginine methyl ester) [27-29].

However, further studies showed, that NO concentration does not affect hyperdynamic circulation and the severity of liver damage in cirrhosis patients. Thus, methylene blue has improved arterial oxygenation, but only temporarily, while L-NAME had no effect on gas exchange in many patients with HPS [30].

There can be no doubt that the increased production of nitric oxide is an important factor underlying the molecular mechanisms that cause pulmonary vascular dilatations in the hepatopulmonary syndrome, but future research is required into the mediators of pulmonary vascular change in HPS in the hope of identifying a target for a potential medical therapy.

Thus, in rats with different models of hepatopulmonary syndrome activation of nitroxydergic process by a significant increase in total concentration of $\text{NO}_2^- + \text{NO}_3^-$ and total NO synthases activity in blood serum and lung tissue supernatant has been established. Herewith a more pronounced intensification of nitroxydergic processes was observed in rats on the 28th day after the common bile duct ligation. Comparing the total concentration of $\text{NO}_2^- + \text{NO}_3^-$ in blood serum and lung tissue supernatant in both models of hepatopulmonary syndrome, we have found the synchronous development of nitroxydergic processes on systemic and local levels and predominance of nitrogen (II) oxide synthesis in lungs.
Study limitation. There are some limitations in the present study. Firstly, this research was conducted only on a small size of rat population. It is therefore essential to validate our findings with greater sample sizes to determine the features of nitroxydergic processes. Secondly, the present study investigated only 2 experimental hepatopulmonary syndrome models, which do not reflect all real causes in patients.

Conflict of interest statement. The authors stated that they have no conflicts of interest regarding the publication of this article.


References


Received 15.05.2018