

REVIEW

UDC 578.834.1+616.379-008.64

doi: <https://doi.org/10.15407/ubj95.02.005>

IMMUNOLOGICAL MECHANISMS OF INCREASED SUSCEPTIBILITY TO COVID-19 DISEASE AND ITS SEVERE COURSE IN PATIENTS WITH DIABETES MELLITUS TYPE 2 AND OBESITY

K. P. ZAK¹, M. D. TRONKO¹, S. V. KOMISARENKO²✉

¹V. P. Komisarenko Institute of Endocrinology and Metabolism,
National Academy of Medical Sciences of Ukraine, Kyiv;

²Palladin Institute of Biochemistry, National Academy of Sciences of Ukraine, Kyiv;
✉e-mail: svk@biochem.kiev.ua

Received: 28 April 2023; Revised: 28 May 2023; Accepted: 05 June 2023

In this review, we analyze and summarize literature data and the results of our own research related to the immunity status of patients with type 2 diabetes mellitus (T2D) and those T2D patients who were infected with the SARS-CoV-2 virus. It was shown that in the blood plasma of T2D patients, especially those with elevated BMI, the level and ultrastructure of the main cellular components of natural immunity - neutrophils and monocytes - were affected accompanied by high levels of proinflammatory cytokines (IL-1 β , IL-6, IL-17 and TNF- α). It was suggested that the increased susceptibility of T2D patients to SARS-CoV-2 infection is primarily due to a weakening of the innate immune defense against pathogens, whereas in T2D patients who have COVID-19, adaptive T-cell immunity disorders accompanied by a cytokine storm prevail. It was concluded that hyperinflammation in T2D+COVID19 patients is the result of enhancement of already existing before SARS-CoV-2 infection T2D-caused disorders of innate and adaptive immunity, in the mechanism of which cytokines and chemokines play a significant role.

Key words: type 2 diabetes mellitus, COVID-19, cytokines, T-lymphocytes, neutrophils, innate and adaptive immunity.

Until very recently, the world experienced the greatest tragedy of our time – the coronavirus COVID-19 pandemic, which took the lives of millions on our planet. Thus, according to the WHO data, the number of COVID-19 patients global by May 2023 reached about 766 million people including 6.9 million deaths [1]. Yet it has been firmly established that old age, obesity and diabetes mellitus (DM) are the main risk factors for SARS-CoV-2 infection, the more severe its clinical course and increased mortality, especially upon acute respiratory distress syndrome (ARDS), as well as increased level of hyperglycemia and insulin resistance (IR) [2-4].

Analysis of the data obtained from the case histories of thousands of COVID-19 patients ad-

mitted to the Central Hospital of New York (USA) showed that 34% of these patients were simultaneously diagnosed with type 2 diabetes (T2D) [5]. A high rate (39.6%) of T2D among patients suffering from COVID-19 was also observed at Massachusetts General Hospital (USA) [6]. A significant percentage (23.3%) of T2D in patients with morbid COVID-19 was also reported in Lombardy (Italy) [7]. Moreover, a significantly higher mortality rate was observed in patients with COVID-19+T2D than patients with COVID-19 without T2D. According to some data, this ratio was 42.8% versus 21.7% ($P < 0.001$) [7], and to others - 20.3% versus 10.9% ($P < 0.01$), respectively [8].

A more severe clinical course of the disease and increased mortality rate were also observed in

obese ($\text{BMI} > 30 \text{ kg/m}^2$) COVID-19 patients with T2D compared to normal patients ($\text{BMI} \leq 25 \text{ kg/m}^2$) [9, 10].

To answer the questions – why patients with T2D are more prone to SARS-CoV-2 infection and why in T2D patients who have already caught COVID-19, especially those with obesity, the disease is more severe with a higher mortality rate – more detailed information about the immune defense system in the T2D patients before their infection with COVID-19 is required.

For many years, we have studied natural and T-cell immunity in newly diagnosed untreated T2D patients with a range of body weights [11-16]. The present work summarizes and analyzes these numerous studies, as well as current literature data on the immunity of individuals with prediabetes ($\text{HbA1c} \geq 5.7\text{-}6.4\%$) and T2D ($\text{HbA1c} > 6.5\%$) in comparison with the results of published studies in patients with COVID-19 of varying severity [17-28], which could be important for understanding the causes and immune mechanisms that make COVID-19 more severe in T2D patients as well as for developing more targeted treatments.

Natural and T-cell immunity in T2D patients with a range of body weight

According to modern concepts, T2D is a chronic low-gradient systemic immune-based inflammation [29-32]. Thus, most of the classic inflammatory biomarkers were found in patients with T2D, namely: leukocytosis, neutrophilia, monocytosis, increased level of C-reactive protein, NLR index, ferritin, D-dimer [33-35], as well as high levels of pro-inflammatory cytokines (IL-1 β , IL-6, IL-17, TNF α , etc.) [14, 34, 36-43].

A statistically significant increase in similar inflammatory biomarkers, but to a lesser extent, was also observed in normoglycemic individuals with morbid obesity [44-49].

It should be noted that the vast majority of patients with T2D (almost 80% of cases) tend to suffer from comorbid obesity [50, 51], which is now recognized to be a low-gradient inflammation of adipose tissue (AT), similar to T2D [52]. Furthermore, the number of overweight and obese people is steadily increasing. About 2 billion people worldwide are obese, that is, every eleventh person on our planet has obesity [53]. It was also discovered that in many countries [54], including Ukraine [55], the curve of the annual global increase in the number of

obese people and the curve of the annual increase in the number of people with T2D run in parallel. According to the IDF data, the number of T2D patients worldwide has quadrupled over the past 25 years and was 537 million people for 2021 and is predicted to rise to 643 million by 2030 [56]. It was also found that each increase in body mass index (BMI) by 5 kg/m^2 increases mortality among T2D patients by 30% [9, 57].

Although obesity is considered one of the main causes of insulin resistance (IR) resulting in the development of T2D in most individuals via the route $\text{AT} \rightarrow \text{IR} \rightarrow \text{PD} \rightarrow \text{T2D}$, it is still unclear whether obesity is the primary cause of T2D or IR.

The age of the patient is also essential in the onset of type 2 diabetes. Thus, one in five people over 65 was diagnosed as having T2D [56].

Our study of the PB leukocyte composition showed [15], that untreated newly diagnosed T2D patients aged 40-65 years had statistically significant leukocytosis on account of an increase in the absolute number of neutrophils and monocytes. The most pronounced segmented neutrophilia (without “left shift”) (Fig. 1) was observed in overweight (especially obese) T2D patients, which correlated with BMI values ($35\text{-}37 \text{ kg/m}^2$). Similar changes were observed in the PB monocyte count, predominantly in obese women (Fig. 1) [58].

At the same time, the absolute number of lymphocytes was slightly but reliably decreased [15], and in some severe patients, especially women, the most pronounced lymphocytopenia was observed. Moreover, the NLR index, an important inflammatory biomarker [59], was significantly increased. The results of our study are fully consistent with the data obtained in the research conducted by immunological centers in various European countries during which a massive number of T2D patients were examined [31, 35].

Our own research [15] also showed that the degree of leukocytosis, neutrophilia, monocytosis and NLR index in T2D patients greatly depended on body weight. Thus, the comparison of leukocyte count in PB of normoglycemic normal weight individuals ($\text{BMI} \leq 25.5 \text{ kg/m}^2$) with normal weight T2D patients ($\text{BMI} \leq 25.5 \text{ kg/m}^2$) revealed a significant increase in total leukocyte count by 11.5%, in neutrophil count by 20.7%, and in monocyte count by 11.1%, NLR index by 19.4%, whereas in obese T2D patients ($\text{BMI} > 35.5 \text{ kg/m}^2$) lymphocyte count was higher by 67.3%, neutrophil count - higher by 93%,

monocyte count – higher by 97%, and NLR index – higher by 54.5% compared to normoglycemic individuals (Fig. 1).

In the normoglycemic control group, similar differences in PB counts between normal weight and obese individuals were detected, but the differences were less pronounced than in T2D patients.

It was revealed, using electron microscopy, that the increased neutrophil count in PB of T2D patients, especially with concomitant obesity, was accompanied by significant changes in neutrophil ultrastructure, indicating a disorder in the functional activity of this type of leukocytes [13].

The PB neutrophils of T2D patients, in comparison with those of normoglycemic individuals, contain a greater number of destructive primary (azurophilic) granules with matrix residues (phagolysis) and many vacuoles (lysosomes) in the cytoplasm (Fig. 2). Neutrophils with an altered structure

of mitochondria and nuclei with an increased number of appendages are more common. The observed changes in neutrophil ultrastructure upon T2D were also confirmed by other methods for studying such cells upon diabetes: phagocytosis, ketosis (extracellular traps), cytotoxicity, which also demonstrated a decrease in the protective anti-inflammatory activity of neutrophils in diabetes [60, 61]. Increased vacuolization of neutrophil cytoplasm, which is a sign of increased secretory activity of cells, is consistent with published data that this type of leukocytes is responsible for increasing production of some proinflammatory cytokines (IL-1 β , IL-6) and chemokines (IL-8) upon T2D [28, 62, 63].

Electron microscopy analysis of leukocytes in T2D patients also revealed pronounced submicroscopic changes in another integral cellular component of natural immunity – monocytes/macrophages. PB monocytes in T2D patients, compared to those

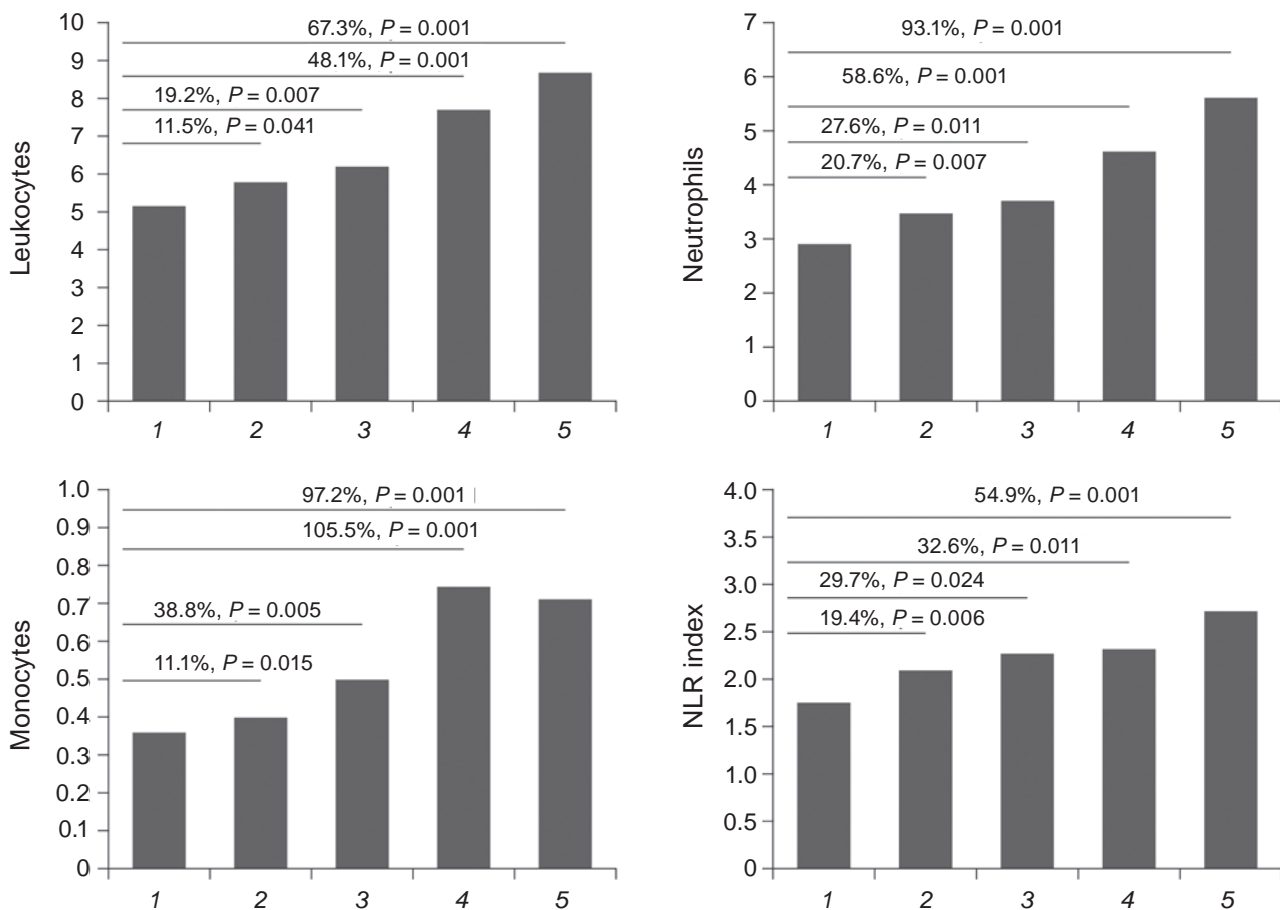


Fig. 1. Total leukocyte count ($10^9/l$), absolute neutrophil count, monocyte count and NLR index in newly diagnosed T2D patients in relation to BMI values: 1 – healthy individuals (BMI < 25.5 kg/m²); 2 – T2D patients (BMI < 25.5 kg/m²); 3 – T2D patients (BMI 25.5 to 29.9 kg/m²); 4 – T2D patients (BMI 30 to 34.9 kg/m²); 5 – T2D patients (BMI > 35 kg/m²) [15]

in normoglycemic individuals, contain a significantly smaller number of specific granules and an increased number of pinocytotic vacuoles and vesicles, involved in the transport and secretory activity of macrophage inflammatory cytokines [13]. Particularly significant changes in the monocyte submicroscopic structure were observed in T2D patients with concomitant obesity.

Thus, our data on pronounced changes in the number and submicroscopic structure of neutrophils and monocytes, essential cellular elements of natural immunity, in T2D patients suggest a significant weakening of the innate (hereditary) antibacterial defense system in this disease.

In studying the number of T-lymphocytes of various immunophenotypes (CD3+T, CD4+T, CD8+T, CD20+ and CD56+ cells) in PB of newly diagnosed T2D patients by flow cytometry using a FACS-tar plus laser flow cytometer and a panel of labeled monoclonal antibodies for cluster differentiation (CD), a significant increase in the total number of T-lymphocytes (CD3+T-cells) and their major subclasses: CD8+T and especially CD4+T-cells were observed, especially in women (Fig. 3).

Some reports noted that a significant increase in the number of PB CD4+T-cells and their recruitment to the AT was observed mainly in obese women with T2D [64, 65] and less pronounced in normoglycemic ones [47, 66, 67]. Moreover, it was found that CD4+T-cells affecting the Th17 cell production are the main producers of pro-inflammatory cytokine IL-17 [47]. However, there are reported data on a decrease in the regulatory fraction of CD4+, CD25+Fox3 lymphocytes in PB of obese patients both with and without T2D [68].

Using electron microscopy analysis of an enriched concentrate of CD4+T-cells isolated by flow cytometry we revealed that most lymphocytes of this subpopulation contained a specific organelle in the cytoplasm, the Goll bodies, which is a reliable cytoplasmic biomarker of CD4+T-cells [69, 70]. The Goll body cluster (GB) of CD4+ T-cells consists of a large rounded granule surrounded by smaller electron-dense satellite granules (Fig. 4, A and B). The GB cluster is reported to play an important role in the CD4+T cells secretory function, including the production of IL-6 and IL-17 cytokines [47, 71]. In T2D patients, especially those with morbid obesity (BMI > 30 kg/m²), significant changes in the submicroscopic structure of the GB cluster (arrows) in the form of an increased number of smaller electron-

dense satellite granules and an increased number of transport vesicles and vacuoles were detected (Fig. 4, A, B). This confirmed the data on significant changes in CD4+T-cell function in T2D patients [13, 72].

It is now well established that cytokines, hormone-like low-molecular-weight proteins or polypeptides that regulate most vital physiological and pathophysiological processes in the body, play a key role in the pathogenesis of inflammatory diseases, including T2D. Cytokines exhibiting apocrine, paracrine and distant (i.e., endocrine) effects are involved in cell signaling in the immune response and protection against various pathogens and stress and are also crucial in the regulation of the cardiovascular system, renal function, hematopoiesis, obesity, carcinogenesis, atherosclerosis, aging, as well as T2D [13, 27, 73].

The central role in the development of IR, PD and clinically evident T2D is attributed to the main proinflammatory cytokines, such as IL-1 β , IL-6, IL-17 and TNF α , considered classic biomarkers of inflammation [27, 28, 36, 74].

Patients with T2D were shown to have elevated levels of the proinflammatory macrophage IL-1 β (Fig. 5, A) in both the preclinical and clinical stages of the disease [14, 75]. Significant increases in IL-1 β M1 macrophage levels in the AT of T2D patients in the postprandial stage were also reported [75]. Especially high levels of IL-1 β were observed in obese T2D patients (BMI = 35-40 kg/m²) [76]. There was also evidence of a significant decrease in the level of IL-1 β in PB of normoglycemic obese individuals following their long-term diet and weight loss [77]. It was reported that the blockage of IL-1 receptor antagonist (IL-1Ra), which can neutralize the effect of IL-1, led to a sharp decrease in IL-1 concentrations in PB and a reduction in the incidence of T2D in patients with high IL-1 levels [78-80].

Numerous prospective studies such as widely known international projects MONICA, EPIC [27, 36, 74, 81, 83] and our own research [13, 14] conducted on patients with newly diagnosed and chronic T2D showed that patients with T2D are characterized mostly by an increased level of the proinflammatory cytokine IL-6 in PB compared to healthy individuals. The meta-analysis of published data fully confirmed the above data [74]. Moreover, elevated levels of IL-6 can be detected long before the development of T2D and, therefore, can be used as a reliable biomarker for predicting the risk of the disease [81]. It was also demonstrated that the de-

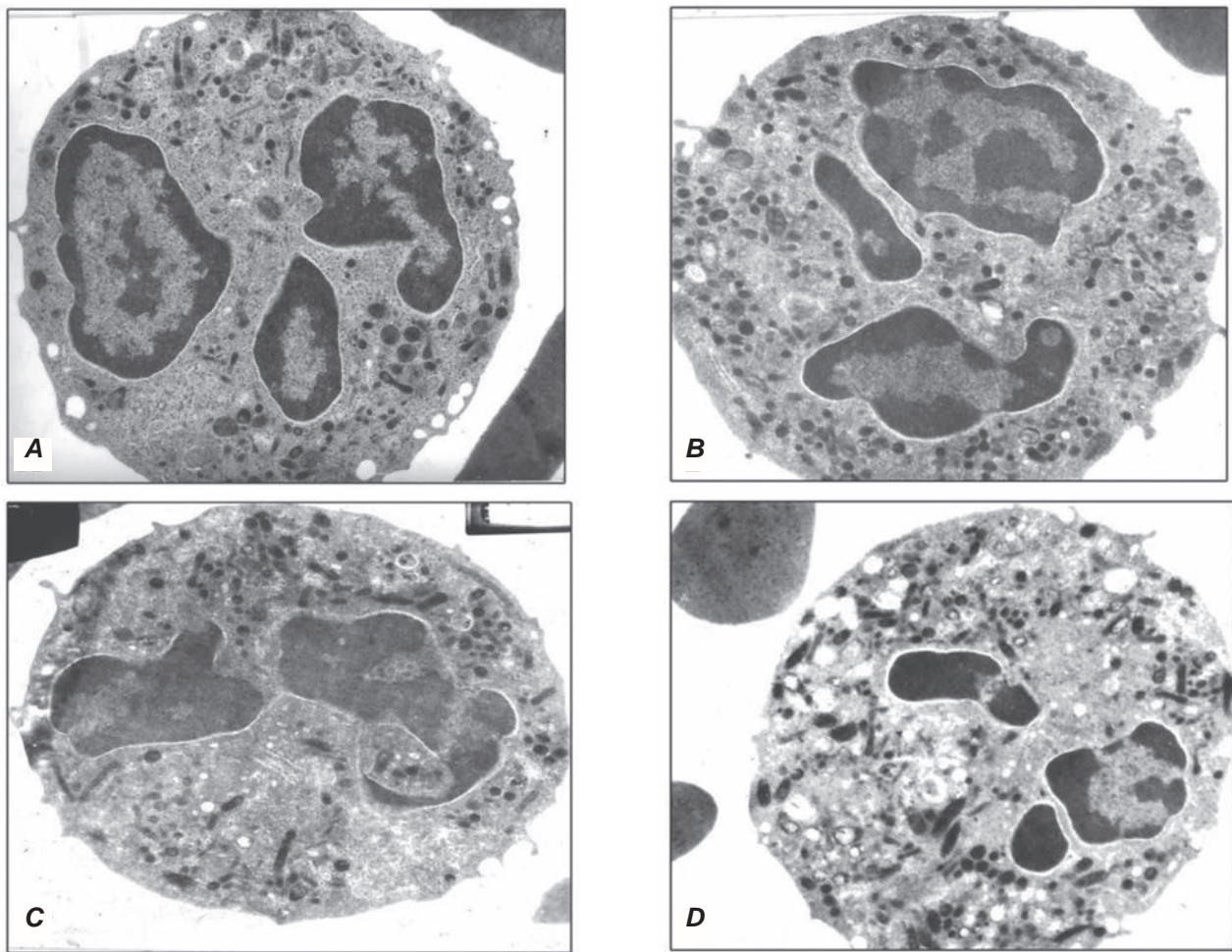


Fig. 2. Segmentonuclear neutrophils in PB: **A** – healthy individual (control) with normal BMI (23.5 kg/m²); **B** – a newly diagnosed T2D patient with normal BMI (22.8 kg/m²); **C** – individual with metabolic syndrome/obesity (BMI 35.0 kg/m²); **D** – T2D patient with metabolic syndrome/obesity (BMI 37.0 kg/m²), $\times 9000$ [42]

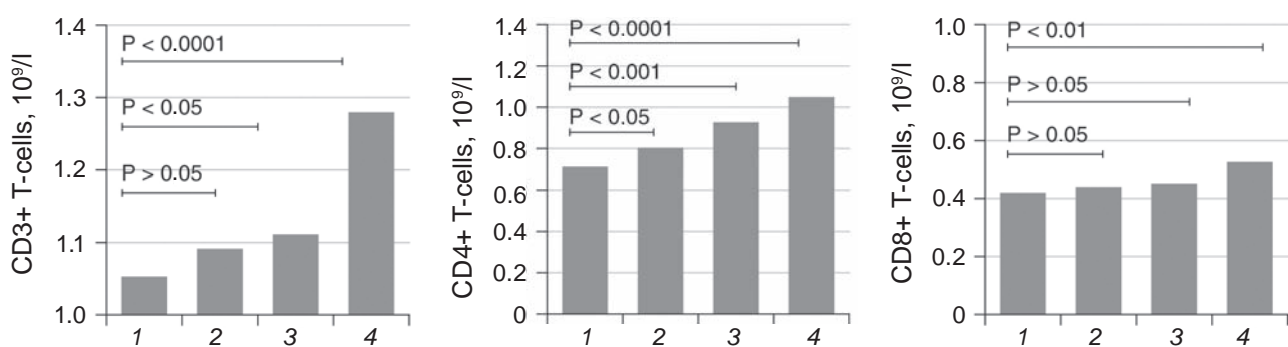


Fig. 3. Absolute counts (10⁹/l) of CD3+ T-, CD4+ T- and CD8+ T-cells in 1 – normoglycemic individuals with BMI < 25 kg/m² and T2D patients with a range of body weights: 2 – BMI < 25 kg/m²; 3 – BMI 25-30 kg/m²; 4 – BMI > 30 kg/m² [16]

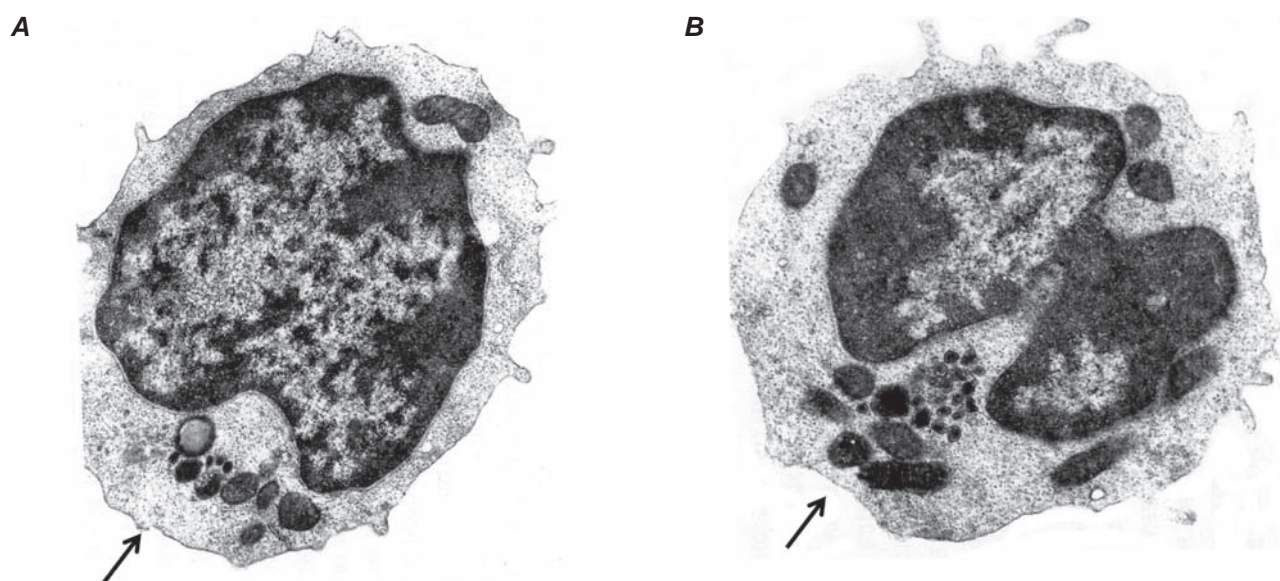


Fig. 4. Ultrastructure of CD4⁺ T-cells containing Golgi bodies (arrows). **A** – healthy individuals (BMI < 25 kg/m²); **B** – T2D patients with obesity [13]

gree of the IL-6 increase in the PB of diabetic patients depended largely on the value of body mass index (BMI), i.e., concomitant obesity (Fig. 5, *B*) [13, 83, 84].

In T2D patients, a significant increase in the content of recently discovered proinflammatory cytokine IL-17A, a highly active immunomodulator of natural and T-cell immunity, was found [47, 85, 86]. Moreover, it was shown that in patients with IR and obesity, the level of IL-17 generation in AT increases almost 8-10-fold [47]. We also revealed a similar increase in IL-17 level (Fig. 5, *B*) in patients with newly diagnosed T2D compared to normoglycemic individuals.

Another characteristic feature of T2D patients was found to be a significant increase in the proinflammatory cytokine TNF α , secreted mainly by the AT cells [13, 14, 84, 87, 88]. It was shown that elevated plasma levels of TNF α in normoglycemic individuals may precede clinically diagnosed T2D and can serve as a marker for identifying a risk group among them [36]. A particularly pronounced increase in TNF α level in PB was observed in T2D patients with concomitant obesity [45, 84, 89-91].

Our studies showed that the degree of increase in all proinflammatory cytokines in T2D patients is largely correlated with the value of body weight/obesity (Fig. 5), i.e. the higher the patient's BMI, the higher PB cytokine levels [12, 13]. These findings suggest that since most T2D patients usually have an elevated BMI, hypercytokinemia is not only the

result of the pathogenesis of T2D itself but is also largely because of the concomitant excess body weight. Further research is needed to support this conclusion.

The role of anti-inflammatory cytokines (IL-4, IL-10, IL-13, IL-38, TGF β) in T2D pathogenesis has been discussed only in a few ambiguous publications [92]. Among them, the most well-known are studies of the cytokine contents of IL-4 and IL-10 in the PB and AT of T2D patients. These types of cytokines are characterized by high pluripotency and gene polymorphism [93-95].

The pleiotropic anti-inflammatory cytokine IL-4, produced mainly by Th-2 and mastocytes, exhibits the ability to bind to specific cell receptors and is involved in the signaling cascade resulting in a direct protective effect of pancreatic islets against pathogen damage. Incubation of IL-4 with adult pancreatic islets prevents apoptosis caused by the “cocktail” of proinflammatory cytokines IL-1 β + TNF α + IFN γ [96].

Some research investigating the IL-4 level in PB of adult T2D patients with normal BMI reported conflicting data. Some studies [97, 98] found decreased levels of IL-4 in PB, while others observed elevated or near-normal levels [99, 100]. A particularly significant increase in IL-4 level was observed in African-American women with high HbA1c levels [91]. The authors attributed the differences in IL-4 levels in PB of T2D patients across continents to genotype polymorphism [101].

Fig. 5. The median levels of IL-1 β (A), IL-6 (B), IL-17 (C), and TNF α (D) in the PB of normoglycemic individuals (control, BMI < 25 kg/m²); T2D patients with BMI < 25.0 kg/m²; normoglycemic individuals with metabolic syndrome/obesity (BMI > 30 kg/m²); and T2D patients with concomitant metabolic syndrome/obesity (BMI > 30 kg/m²)

the production of IL-6 and TNF α . In individuals with IR as well as T2D patients, a decrease in the IL-10 production and PB plasma level, especially pronounced in metabolic syndrome and obesity [103, 104], were observed [33, 103]. However, there

are data that in T2D patients, especially Brazilians and Italians, a slight increase in PB IL-10 levels were registered [83, 105].

The paradoxical action of the anti-inflammatory cytokines IL-4 and IL-10 in T2D is believed to be a protective compensatory response of the body to reduce the high level of pro-inflammatory cytokines so as to maintain a normal balance of pro- and anti-inflammatory cytokines [83, 92].

An important role in T2D pathogenesis is also performed by numerous chemokines. The level of CCL-2 in PB of T2D patients was found to be significantly increased [106, 107]. There was demonstrated a direct correlation between the CCL-4 level in PB and IR: CCL-4 deficiency significantly improves IR [108]. A marked increase in the level of chemokine CCL-5 in T2D patients was shown [109]. The RANTES/CCL-5 chemokine was reported to contribute to the IR development and was detected in MS and T2D [107].

A marked increase in the chemoattractant CXCL-8 (IL-8) upon T2D was also shown. It was established that this chemokine precedes the development of T2D [110]. In an *in vitro* study of isolated Langerhans islets in T2D patients, it was revealed that they secreted 33.5-fold more IP-10/CXCL-10 chemokine. A significant increase in CX3CL-1 in PB of T2D patients was observed [111]. These findings are considered to support the hypothesis of a significant role of the CX3CL-1 CX3CP-1 system in relation to AT inflammation, IR, and T2D.

For the first time, we showed that the chemokine IL-16/CXCL-16 is also actively involved in T2D pathogenesis [11], and it was confirmed by foreign researchers [112].

In conclusion it is worth noting that the above described changes in immunity indices were also observed in individuals with IR and the metabolic syndrome (prediabetes).

Natural and T-cell immunity in SARS-CoV-2-infected T2D patients

COVID-19 is an acute severe immune-mediated inflammatory disease caused by the SARS-CoV-2 virus or its mutants.

The mechanism of SARS-CoV-2 virus penetration into uninfected human cells is rather complicated and has not yet been sufficiently studied. The hypothesis based on studies [113-116] suggests that the key participants of SARS-CoV-2 virus penetration into epithelial cells are Spike pro-

teins, which cover the virus spike surface, S-protein with glucose-regulating protein and receptor to angiotensin-converting enzyme (ACE).

At the same time, it was discovered that the proinflammatory transcription factor NF- κ B and the signal transducer JNK including the activator of transcription 3 (STAT3) also play a critical role in the mechanism of SARS-CoV-2 virus entry [10].

Among COVID-19 patients, as it has already been noted, considerable percentages were patients with morbid T2D. In these COVID-19 patients, as in COVID-19 patients without diabetes, high levels of inflammatory biomarkers such as neutrophilia [13, 15], C-reactive protein [18, 20, 28, 117], ferritin [28, 119], D-dimer [20, 118], NLR index [28, 63], and increased proinflammatory cytokines [17, 19, 20] were also commonly observed.

Changes in natural and adaptive immunity parameters in COVID-19 patients were largely similar but more pronounced than in T2D patients, especially with concomitant obesity [2, 3].

In the hematological study of COVID-19 patients, pronounced changes in the leukocytic composition of PC were shown to be similar to those in T2D, namely by an increase in the total number of leukocytes [21, 23, 28, 63] the absolute number of neutrophils and partly monocytes [21, 23, 63, 120] with a decrease in the absolute number of lymphocytes [17, 19, 20, 120].

Significant neutrophilia, greater than that in T2D, was found in COVID-19 patients with existing complications, especially upon acute respiratory distress syndrome (ARDS) [2, 9, 10, 21, 23, 25], as well as concomitant T2D and obesity (BMI > 30 kg/m²) [5-7, 10, 121, 122]. It is also believed that hyper neutrophilia in COVID-19 may be the most reliable biomarker of ARDS risk [22, 23].

At the same time, it should be noted that neutrophils are important cellular components involved in innate immunity [69, 123] and serve as producers of a range of major proinflammatory cytokines and chemokines [28, 63].

COVID-19 patients, compared to patients with T2D, are also characterized by marked lymphocytopenia, especially high in acute respiratory distress syndrome (ARDS), which largely results from the interaction between AP-1 located on SARS-CoV-2 and renin-angiotensin system receptors located on pneumocytes. Thus, very high lymphocytopenia can be an indicator of ARDS and high mortality risk [17-20, 23, 28, 120].

The human lymphocyte population comprises many subpopulations, often characterized by different functional activities, and so the question arises: which subpopulation is responsible for lymphocytopenia? There are ambiguous publications on the study of lymphocyte immunophenotype in patients infected with the SARS-CoV-2 virus that have not yet given a clear insight into the T-cell immunity status in this disease [114]. Thus, according to some data [20, 120], in patients with severe COVID-19, a decrease in the total number of T-lymphocytes (CD3+ T-cells) was due to a decrease in both of main lymphocyte subclasses, i.e. CD4+T- and CD8+ T-cells. According to other data [25], lymphocytopenia in patients with the severe form of COVID-19 was mainly due to a decrease in the CD4+ T-cell subpopulation, while the CD8+ T-cell subpopulation decreased to a much lesser degree. Moreover, the decrease in the number of CD4+ T-cells occurred mainly due to a decrease in the subpopulation of memory CD45 Ro+ cells and regulatory CD25+ CD127 low+ cells, while the number of naïve CD45 Ro- cells increased. According to these authors, the high number of naïve memory cells in patients with moderate illness indicates that the ratio of different immune cells may change during the COVID-19 disease.

Some studies revealed a significant decrease in all T-lymphocyte subpopulations (CD3+T-, CD4+T- and CD8+T-cells) in patients with COVID-19, which correlated with disease severity and mortality rate. Based on their study, the authors concluded that determining the counts of various T-lymphocyte subpopulations is a reliable biomarker for early diagnosis of COVID-19 and potential mortality risk [24].

In COVID-19 patients with moderate and especially severe disease, as in T2D patients, there were detected simultaneously both neutrophilia and lymphocytopenia that leads to a significant increase in the NLR index [28, 63, 124, 125], also called the “death index” [59, 128].

Also, it was shown [2, 26] that the increased levels of monocytes/macrophages in COVID-19 patients contribute greatly to ARDS pathogenesis through the hypersecretion of cytokines IL-1 β and IL-6.

Summarizing our studies on significant disorders in the content and ultrastructure of PB neutrophils and monocytes, the main cellular components of natural immunity, in T2D patients, we can conclude that the increased susceptibility of T2D patients to SARS-CoV-2 infection is the result of the

weakening of natural immune protection probably driven, in large part, by genetic predisposition.

Still, one of the most marked immune dysfunctions in T2D patients, especially those with developed COVID-19, is a sharp increase in PB content of various proinflammatory cytokines and chemokines, an event known as cytokine storm syndrome [17, 19, 20, 116].

About 20 different types of cytokines have already been studied in COVID-19 patients. The most studied are proinflammatory cytokines (IL-1 β , IL-6, IL-7, IL-17, IL-18, TNF α), anti-inflammatory (IL-4, IL-10), immunomodulators (IL-2, IFN α and IFN γ), regulators of blood formation (CRP-H) and chemokines (CCL-2, CCL-4, CCL-5, CCL-8, CCL-10).

The degree of changes in the content of different types of cytokines in SARS-CoV-2-infected patients depends largely on the stage of the disease, its severity, the medical care quality and intensity, the complications developed, especially lung injury (ARDS), and comorbid morbidities, especially obesity and T2D [17, 22, 26, 116, 127].

The most pronounced changes in proinflammatory cytokines in COVID-19 patients, as well as in T2D patients, were found in cytokines IL-1 β , IL-6, TNF α , and IL-17.

Thus, some publications reported that in COVID-19 patients, elevated levels of the proinflammatory macrophage cytokine IL-1 β were detected [26, 128]. According to these data, in severe COVID-19 patients, the cytokine IL-1 β activates and promotes the secretion of inflammasomes, a complex of intracellular multiproteins, leading to lytic cell death (pyroptosis) [128].

According to the majority of researchers [18, 17, 19, 20, 114], the highest content in PB in COVID-19, among different types of proinflammatory cytokines, was observed in IL-6, also called the “king of cytokines”. At critical ICU and ARDS occurrence, the level of IL-6 in PB plasma in COVID-19 patients can reach 150 pg/ml or more (at norm: 1.4 ± 1.2 pg/ml), which is associated with the worst scenario, that is an increase in mortality rate [112]. In this regard, it was suggested that IL-6 should be considered the most accurate biomarker of COVID-19 disease severity and mortality risk [118, 120].

In COVID-19 patients, as well as in T2D patients, a significant increase in the content of TNF α , another major proinflammatory cytokine, secreted mainly by AT macrophages, was found [17, 19, 20, 118]. A particularly high level of this cytokine was

observed in obese women with COVID-19. Moreover, the high level of TNF α significantly decreased after therapy with metformin, which simultaneously lowered both hyperglycemia and BMI value [129].

There are also few reports on an increase in the proinflammatory cytokine IL-17 in COVID-19, similar to that in T2D [112].

Some unexpected data demonstrated a significant increase in the anti-inflammatory cytokines IL-4 and IL-10 in SARS-CoV-2-infected patients in contrast to T2D patients [17, 20, 25, 118].

Given this paradox, it was suggested that the increased level of anti-inflammatory cytokines IL-4 and, especially, IL-10 in COVID-19 patients resulted from the activation of protective compensatory mechanisms to restore the balance between pro- and anti-inflammatory cytokines [83, 91].

Important roles in protecting the body against SARS-CoV-2 were shown to play also by the immune-regulating cytokine IFN γ , which exerts a modulating effect and changes immunotolerance upon infection [117, 130], as well as the colony-stimulating cytokine CSF-H, which controls neutrophilopoiesis [17, 25, 118].

In COVID-19 patients, a significant increase in PB chemokines, such as CCL2, CCL5, CXCL8, CXCL10, CXCL16, was also detected that may explain the mechanism of mass migration of various types of immunological cells into inflammation focus [17, 19, 112, 118].

Until recently, the possibility of a direct inhibitory effect of the SARS-CoV-2 virus on insulin production by pancreatic beta cells remained subject to debate. Based on the analysis of case histories, a number of practitioners suggested that the SARS-CoV-2 virus could directly affect the mechanisms involved in the onset of diabetes in COVID-19 patients [122, 131, 132]. However, this assumption has been met with reasonable criticism [133].

Thus, using molecular and ultracytochemical methods, pronounced replication, differentiation, and degradation of beta cells in the pancreas, taken at biopsies from COVID-19 patients, were first described [134].

These data were confirmed by the study [135], showing that in COVID-19 patients, high susceptibility of beta cells to SARS-CoV-2 and secretion of cytokines and chemokines along with pronounced beta cell transdifferentiation, their degranulation and low insulin expression were observed [135].

A retrospective analysis of data obtained in a study of about 40,000 COVID-19 patients showed

that the occurrence of primary T2D a short time after infection with SARS-CoV-2 was found to be 12.3 people per 1,000 patients [136].

T2D combined with overweight/obesity is considered one of the main risk factors for COVID-19 development [2, 6, 10]. Moreover, symbiosis of all three types of inflammation (COVID-19+T2D+obesity) resulted in the most severe course of COVID-19 disease and dramatically increased mortality [7, 9, 10].

In recent years, important new data have also been published that support the concept that T2D and obesity are chronic low-grade inflammatory diseases that give clues to the mechanism of effectiveness of present-day COVID-19 therapy. First of all, it was found that a number of widely-used hypoglycemic antidiabetic drugs such as insulin, glycogen-like polypeptide agonists, thiazolidinediones exhibit a pronounced anti-inflammatory effect, leading to a decrease in the elevated levels of proinflammatory cytokines (IL-1 β , IL-6, TNF α) [32]. A particularly significant anti-inflammatory and immunosuppressive effect, as well as a positive therapeutic effect, and a decrease in the mortality rate among T2D patients infected with SARS-CoV-2 were shown for the antidiabetic drug metformin [129, 137, 138]. This can be attributed to the ability of metformin, along with its hypoglycemic effect, to normalize the levels of proinflammatory cytokines and other inflammatory biomarkers, as well as to reduce BMI [140]. It was very surprising discovery that the synthetic anti-inflammatory corticosteroids dexamethasone, prednisolone, and hydrocortisone possessed a particularly high positive therapeutic effect in severe COVID-19 patients. It was reported that the treatment of severe COVID-19 patients with massive doses of dexamethasone resulted in a significant reduction in the mortality rate, from approximately 40% (placebo) to 20% [140, 141].

Conclusion. The currently available data on the status of natural and adaptive immunity in patients with COVID-19, as well as T2D, accompanied by severe inflammation show that their immune parameters, in comparative terms, are very similar in many ways. This view was confirmed by increased levels of almost all major inflammation biomarkers such as neutrophilia, monocytosis, NLR inflammatory index, C-reactive protein, ferritin, D-dimer and proinflammatory cytokines in both diseases.

For T2D and COVID-19 patients, a significant increase in the absolute number of neutrophils and monocytes, the main cellular components of natural

immunity, is very common. Our study of neutrophils and monocytes in T2D patients using electron microscopy revealed pronounced submicroscopic changes in their cytoplasm in the form of degranulation of azurophilic granules and cytoplasm vacuolization. This indicates a significant decline in their function, i.e., the earliest primary stage of the immune defense against pathogens. The findings give a reasonable explanation for the increased susceptibility of T2D patients to SARS-CoV-2 infection.

In T2D patients with developed COVID-19, there predominates a disorder of adaptive immunity in the form of a significant decrease in the absolute number of lymphocytes in PB determined by a decrease in the number of T-lymphocytes (CD3+T-cells) at the expense of their subclass CD4+T-cells. Moreover, in the cytoplasm of CD4+ T-cells in T2D patients were found significant changes in the ultrastructure of Golgi bodies, which are their cytological markers. The highest lymphocytopenia was observed in COVID-19 patients with concomitant T2D and its frequent comorbidity – obesity (COVID-19 + T2DM + obesity).

The most dramatic changes in the immune state upon both types of the disease (COVID-19 and T2D) were found to be in the increased levels of cytokines in the PB, particularly pro-inflammatory cytokines (IL-1 β , IL-6, IL-17 and TNF α) and chemokines (CCL-2, CCL-4, CCL-5, CXCL-8, CXCL-10, and CX3CL-16) resulting in the development of cytokine storm syndrome, the degree of which in PB is associated with the severity of the clinical course of the disease and a risk factor for increased mortality rate.

Thus, analysis of the literature data and the results of our own studies showed that the increased susceptibility of T2D patients to SARS-CoV-2 infection is mainly attributed to weakening natural immunity, whereas in T2D patients with developed COVID-19, adaptive immunity disorders predominate. Moreover, the extremely severe clinical course and increased mortality rate are considered to be a consequence of summation and potentiation of hyperinflammation and oxidative stress already occurring in T2D resulting from impaired both natural and T-cell immunity, in the mechanism of which cytokines and chemokines play a crucial role.

Conflict of interest. The authors have completed the Unified Conflicts of Interest form at http://ukr-biochemjournal.org/wp-content/uploads/2018/12/coi_disclosure.pdf and declare no conflict of interest.

ІМУНОЛОГІЧНІ МЕХАНІЗМИ ПІДВИЩЕНОЇ СХИЛЬНОСТІ ДО ЗАХВОРЮВАННЯ НА COVID-19 ТА ЙОГО ТЯЖКОГО ПЕРЕБІГУ У ХВОРИХ НА ЦУКРОВИЙ ДІАБЕТ 2 ТИПУ ТА ОЖИРІННЯ

К. П. Зак¹, М. Д. Тронько¹,
С. В. Комісаренко²✉

¹Інститут ендокринології та обміну речовин ім. В. П. Комісаренка НАМН України, Київ;

²Інститут біохімії ім. О. В. Палладіна НАН України, Київ;

✉e-mail: svk@biochem.kiev.ua

Огляд присвячено аналізу літературних даних та результатів власних досліджень авторів щодо стану імунітету у хворих на цукровий діабет типу 2 (ЦД2) та у хворих на ЦД2, інфікованих вірусом SARS-CoV-2. Показано, що у хворих на ЦД2, особливо, з надлишковим індексом маси тіла, порушеними є вміст та ультраструктура основних клітинних елементів природного імунітету – нейтрофілів та моноцитів, що супроводжується підвищеним вмістом прозапальних цитокінів (IL-1 β , IL-6, IL-17 і TNF- α) у плазмі крові. Припускається, що підвищена схильність хворих на ЦД2 до зараження вірусом SARS-CoV-2 обумовлена, перш за все, ослабленням вродженого імунного захисту організму, тоді як у хворих на ЦД2 з розвиненим COVID-19 превалюють порушення адаптивного Т-клітинного імунітету, що супроводжується синдромом “цитокінового шторму”. Зроблено висновок, що гіперзапалення у хворих на ЦД2+COVID-19 є результатом потенціювання вже існуючих порушень вродженого імунітету у хворих на ЦД2 ще до їх зараження SARS-CoV-2, та адаптивного імунітету, у механізмі якого суттєву роль відіграють цитокіни та хемокіни.

Ключові слова: цукровий діабет типу 2, COVID-19, цитокіни, Т-лімфоцити, нейтрофіли, вроджений та адаптивний імунітет.

References

1. WHO Coronavirus (COVID-19) Dashboard <https://covid19.who.int/table>.
2. Mauvais-Jarvis F. Aging, Male Sex, Obesity, and Metabolic Inflammation Create the Perfect Storm for COVID-19. *Diabetes*. 2020; 69(9): 1857-1863.

3. Shin JA, Oh S, Jeong JM. The potential of BEN815 as an anti-inflammatory, antiviral and antioxidant agent for the treatment of COVID-19. *Phytomed Plus*. 2021; 1(4): 100058.
4. Koh HE, van Vliet S, Pietka TA, Meyer GA, Razani B, Laforest R, Gropler RJ, Mittendorfer B. Subcutaneous Adipose Tissue Metabolic Function and Insulin Sensitivity in People With Obesity. *Diabetes*. 2021; 70(10): 2225-2236.
5. Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW, Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW, Barnaby DP, Becker LB, Chelico JD, Cohen SL, Cookingham J, Coppa K, Diefenbach MA, Dominello AJ, Duer-Hefe J, Falzon L, Gitlin J, Hajizadeh N, Harvin TG, Hirschwerk DA, Kim EJ, Kozel ZM, Marrast LM, Mogavero JN, Osorio GA, Qiu M, Zanos TP. Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized With COVID-19 in the New York City Area. *JAMA*. 2020; 323(20): 2052-2059.
6. Seiglie J, Platt J, Cromer SJ, Bunda B, Foulkes AS, Bassett IV, Hsu J, Meigs JB, Leong A, Putman MS, Triant VA, Wexler DJ, Manne-Goehler J. Diabetes as a Risk Factor for Poor Early Outcomes in Patients Hospitalized With COVID-19. *Diabetes Care*. 2020; 43(12): 2938-2944.
7. Mirani M, Favacchio G, Carrone F, Betella N, Biamonte E, Morengi E, Mazziotti G, Lania AG. Impact of Comorbidities and Glycemia at Admission and Dipeptidyl Peptidase 4 Inhibitors in Patients With Type 2 Diabetes With COVID-19: A Case Series From an Academic Hospital in Lombardy, Italy. *Diabetes Care*. 2020; 43(12): 3042-3049.
8. Shi Q, Zhang X, Jiang F, Zhang X, Hu N, Bimu C, Feng J, Yan S, Guan Y, Xu D, He G, Chen C, Xiong X, Liu L, Li H, Tao J, Peng Z, Wang W. Clinical Characteristics and Risk Factors for Mortality of COVID-19 Patients With Diabetes in Wuhan, China: A Two-Center, Retrospective Study. *Diabetes Care*. 2020; 43(7): 1382-1391.
9. Tate J, Knuiman M, Davis WA, Davis TME, Bruce DG. A comparison of obesity indices in relation to mortality in type 2 diabetes: the Fremantle Diabetes Study. *Diabetologia*. 2020; 63(3): 528-536.
10. Shin J, Toyoda S, Nishitani S, Onodera T, Fukuda S, Kita S, Fukuhara A, Shimomura I. SARS-CoV-2 infection impairs the insulin/IGF signaling pathway in the lung, liver, adipose tissue, and pancreatic cells via IRF1. *Metabolism*. 2022; 133: 155236.
11. Zak KP, Kondratskaia IN, Mel'nichenko SV, Popova VV. Circulating interleukin-16 in blood of patients with metabolic syndrome and type 2 diabetes mellitus. *Lik Sprava*. 2007; (5-6): 46-49. (In Russian).
12. Zak KP, Mankovsky BM, Melnichenko SV, Kondratska IM, Popova VV, Sayenko YaA, Semionova TA, Korpacheva-Zynych OV, Tronko KM, Furmanova OV. Immunity in patients with type 2 diabetes mellitus in complex with concomitant metabolic syndrome/obesity. Communication 2. Role of adipocytokines (interleukin-6, tumor necrosis factor alpha, leptin and adiponectin). *Endokrynologiya*. 2013; 18(2): 26-32.
13. Zak KP, Tronko ND, Popova VV, Butenko AK. Diabetes mellitus. Immunity. Cytokines. Kyiv: Knyha plyus, 2015. 485 p.
14. Zak KP, Popova VV, Orlenko VL, Furmanova OV, Tronko ND. Cytokines in the blood of patients with type 2 diabetes mellitus depending on the level of overweight/obesity (literature review and own data). *Int J Endocrinol*. 2021; 17(7): 534-551.
15. Furmanova OV, Zak KP, Popova V, Tronko MD. Blood leukocyte composition and neutrophil to lymphocyte ratio in patients with newly diagnosed type 2 diabetes mellitus depending on the degree of overweight/obesity. *Int J Endocrinol*. 2020; 16(7): 526-533. (In Russian).
16. Furmanova OV, Kulikovska AV, Popova VV, Zak KP, Tronko MD. Immunophenotype of blood lymphocytes in patients with type 2 diabetes and normal body weight and obesity. *Int J Endocrinol*. 2021; 17(2): 108-115. (In Ukrainian).
17. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020; 395(10223): 497-506.
18. Ruan Q, Yang K, Wang W, Jiang L, Song J. Clinical predictors of mortality due to COVID-19

- based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med.* 2020; 46(5): 846-848.
19. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet.* 2020; 395(10229): 1033-1034.
 20. Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, Wang T, Zhang X, Chen H, Yu H, Zhang X, Zhang M, Wu S, Song J, Chen T, Han M, Li S, Luo X, Zhao J, Ning Q. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest.* 2020; 130(5): 2620-2629.
 21. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, Wang B, Xiang H, Cheng Z, Xiong Y, Zhao Y, Li Y, Wang X, Peng Z. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA.* 2020; 323(11): 1061-1069.
 22. Wu C, Chen X, Cai Y, Xia J, Zhou X, Xu S, Huang H, Zhang L, Zhou X, Du C, Zhang Y, Song J, Wang S, Chao Y, Yang Z, Xu J, Zhou X, Chen D, Xiong W, Xu L, Zhou F, Jiang J, Bai C, Zheng J, Song Y. Risk Factors Associated With Acute Respiratory Distress Syndrome and Death in Patients With Coronavirus Disease 2019 Pneumonia in Wuhan, China. *JAMA Intern Med.* 2020; 180(7): 934-943.
 23. Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, Liu L, Shan H, Lei CL, Hui DSC, Du B, Li LJ, Zeng G, Yuen KY, Chen RC, Tang CL, Wang T, Chen PY, Xiang J, Li SY, Wang JL, Liang ZJ, Peng YX, Wei L, Liu Y, Hu YH, Peng P, Wang JM, J Yang Liu 1, Chen Z, Li G, Zheng ZJ, Qiu SQ, Luo J, Ye CJ, Zhu SY, Zhong NS. Clinical Characteristics of Coronavirus Disease 2019 in China. *N Engl J Med.* 2020; 382(18): 1708-1720.
 24. Xu B, Fan CY, Wang AL, Zou YL, Yu YH, He C, Xia WG, Zhang JX, Miao Q. Suppressed T cell-mediated immunity in patients with COVID-19: A clinical retrospective study in Wuhan, China. *J Infect.* 2020; 81(1): e51-e60.
 25. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, Xie C, Ma K, Shang K, Wang W, Tian DS. Dysregulation of Immune Response in Patients With Coronavirus 2019 (COVID-19) in Wuhan, China. *Clin Infect Dis.* 2020; 71(15): 762-768.
 26. McGonagle D, Sharif K, O'Regan A, Bridgewood C. The Role of Cytokines including Interleukin-6 in COVID-19 induced Pneumonia and Macrophage Activation Syndrome-Like Disease. *Autoimmun Rev.* 2020; 19(6): 102537.
 27. Liu BM, Martins TB, Peterson LK, Hill HR. Clinical significance of measuring serum cytokine levels as inflammatory biomarkers in adult and pediatric COVID-19 cases: A review. *Cytokine.* 2021; 142: 155478.
 28. Fernandez-Botran R, Furmanek S, Ambadapoodi RS, Expósito González E, Cahill M, Carrico R, Akca O, Ramírez JA. Association and predictive value of biomarkers with severe outcomes in hospitalized patients with SARS-CoV-2 infection. *Cytokine.* 2022; 149: 155755.
 29. Duncan BB, Schmidt MI, Pankow JS, Ballantyne CM, Couper D, Vigo A, Hoogeveen R, Folsom AR, Heiss G. Low-grade systemic inflammation and the development of type 2 diabetes: the atherosclerosis risk in communities study. *Diabetes.* 2003; 52(7): 1799-1805.
 30. Donath MY. Multiple benefits of targeting inflammation in the treatment of type 2 diabetes. *Diabetologia.* 2016; 59(4): 679-682.
 31. Ratter-Rieck JM, Roden M, Herder C. Diabetes and climate change: current evidence and implications for people with diabetes, clinicians and policy stakeholders. *Diabetologia.* 2023; 66(6): 1003-1015.
 32. Dandona P, Ghanim H. Diabetes, Obesity, COVID-19, Insulin, and Other Antidiabetes Drugs. *Diabetes Care.* 2021; 44(9): 1929-1933.
 33. Blüher M, Fasshauer M, Tönjes A, Kratzsch J, Schön MR, Paschke R. Association of interleukin-6, C-reactive protein, interleukin-10 and adiponectin plasma concentrations with measures of obesity, insulin sensitivity and glucose metabolism. *Exp Clin Endocrinol Diabetes.* 2005; 113(9): 534-537.
 34. Hu FB, Meigs JB, Li TY, Rifai N, Manson JE. Inflammatory markers and risk of developing type 2 diabetes in women. *Diabetes.* 2004; 53(3): 693-700.
 35. Shitole SG, Biggs ML, Reiner AP, Mukamal KJ, Djoussé L, Ix JH, Barzilay JI, Tracy RP, Siscovick D, Kizer JR. Soluble CD14 and CD14 Variants, Other Inflammatory Markers, and Glucose Dysregulation in Older Adults: The Cardiovascular Health Study. *Diabetes Care.* 2019; 42(11): 2075-2082.

36. Spranger J, Kroke A, Möhlig M, Hoffmann K, Bergmann MM, Ristow M, Boeing H, Pfeiffer AFH. Inflammatory cytokines and the risk to develop type 2 diabetes: results of the prospective population-based European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam Study. *Diabetes*. 2003; 52(3): 812-817.
37. Larsen CM, Faulenbach M, Vaag A, Vølund A, Ehses JA, Seifert B, Mandrup-Poulsen T, Donath MY. Interleukin-1-receptor antagonist in type 2 diabetes mellitus. *N Engl J Med*. 2007; 356(15): 1517-1526.
38. Herder C, Peltonen M, Koenig W, Sütters K, Lindström J, Martin S, Ilanne-Parikka P, Eriksson JG, Aunola S, Keinänen-Kiukaanniemi S, Valle T T, Uusitupa M, Kolb H, Tuomilehto J. Anti-inflammatory effect of lifestyle changes in the Finnish Diabetes Prevention Study. *Diabetologia*. 2009; 52(3): 433-442.
39. Wannamethee SG, Papacosta O, Lawlor DA, Whincup PH, Lowe GD, Ebrahim S, Sattar N. Do women exhibit greater differences in established and novel risk factors between diabetes and non-diabetes than men? The British Regional Heart Study and British Women's Heart Health Study. *Diabetologia*. 2012; 55(1): 80-87.
40. Banerjee M, Saxena M. Interleukin-1 (IL-1) family of cytokines: role in type 2 diabetes. *Clin Chim Acta*. 2012; 413(15-16): 1163-1170.
41. Wang X, Bao W, Liu J, Ouyang YY, Wang D, Rong S, Xiao X, Shan ZL, Zhang Y, Yao P, Liu LG. Inflammatory markers and risk of type 2 diabetes: a systematic review and meta-analysis. *Diabetes Care*. 2013; 36(1): 166-175.
42. Zak KP, Mankovsky BM, Kondratska IN, Popova VV, Saenko JA, Lipska OYe, Semionova TA, Afanasyeva VV. Immunity in patients with type 2 diabetes mellitus with concomitant metabolic syndrome/obesity. Communication 1. Composition of blood leukocytes, immunophenotype of lymphocytes, and ultrastructure of neutrophils. *Endokrynologia*. 2013; 18(1): 27-36.
43. Zak KP, Furmanova OV, Popova VV, Sayenko YaA. The content of pro-inflammatory cytokines IL-1 β , IL-6, IL-17A and TNF α in the blood of patients with type 2 diabetes after therapy with metformin. *Ukr Biochem J*. 2020; 92(6): 105-112.
44. Guarino D, Nannipieri M, Iervasi G, Taddei S, Bruno RM. The Role of the Autonomic Nervous System in the Pathophysiology of Obesity. *Front Physiol*. 2017; 8: 665.
45. Cartier A, Lemieux I, Almeras N, et al. Visceral obesity and plasma glucose-insulin homeostasis: contributions of IL-6 and TNF- α . *Diabetologia*. 2007; 50(S1): 270.
46. Kern PA, Ranganathan S, Li C, Wood L, Ranganathan G. Adipose tissue tumor necrosis factor and interleukin-6 expression in human obesity and insulin resistance. *Am J Physiol Endocrinol Metab*. 2001; 280(5): E745-E751.
47. Fabbrini E, Cella M, McCartney SA, Fuchs A, Abumrad NA, Pietka TA, Chen Z, Finck BN, Han DH, Magkos F, Conte C, Bradley D, Fraterrigo G, Eagon JC, Patterson BW, Colonna M, Klein S. Association between specific adipose tissue CD4⁺ T-cell populations and insulin resistance in obese individuals. *Gastroenterology*. 2013; 145(2): 366-374.e1-3.
48. Zak KP, Popova VV. The prediction of type 1 diabetes development and diagnosis of its asymptomatic phase using autoantibodies to human islets of Langerhans long before the onset of the disease. *Int J Endocrinol*. 2016; (7(79)): 11-21. (In Russian).
49. Zak KP, Popova VV, Gruzov MA, Khomenko BM, Afanasyeva VV, Malynovskaya TN, Tronko EN, Saenko YaA, Semionova TA, Kulskovskaya AB. Results of twenty-year studies on immunity at preclinical asymptomatic phase of developing type 1 diabetes in children on the program IPDM: 1. Leukocyte composition and immune phenotype of blood lymphocytes. *Endokrynologia*. 2017; 22(3): 201-210.
50. Lichiardopol R, Popescu LD, Ionescu I, et al. Abdominal obesity in type 1 and type 2 diabetes patients. *Diabetologia*. 2008; 51(Suppl 1): S335.
51. Kim S, Parks CG, DeRoo LA, Chen H, Taylor JA, Cawthon RM, Sandler DP. Obesity and weight gain in adulthood and telomere length. *Cancer Epidemiol Biomarkers Prev*. 2009; 18(3): 816-820.
52. Cox AR, Chernis N, Bader DA, Saha PK, Masschelin PM, Felix JB, Sharp R, Lian Z, Putluri V, Rajapakshe K, Kim KH, Villareal DT, Armamento-Villareal R, Wu H, Coarfa C, Putluri N, Hartig SM. STAT1 Dissociates Adipose Tissue Inflammation From Insulin Sensitivity in Obesity. *Diabetes*. 2020; 69(12): 2630-2641.

53. Kim JY, Bacha F, Tfayli H, Michaliszyn SF, Yousuf S, Arslanian S. Adipose Tissue Insulin Resistance in Youth on the Spectrum From Normal Weight to Obese and From Normal Glucose Tolerance to Impaired Glucose Tolerance to Type 2 Diabetes. *Diabetes Care*. 2019; 42(2): 265-272.
54. Nolan JJ, Færch K. Estimating insulin sensitivity and beta cell function: perspectives from the modern pandemics of obesity and type 2 diabetes. *Diabetologia*. 2012; 55(11): 2863-2867.
55. Tronko ND, Zak KP. Obesity and diabetes mellitus. *Lik Sprava*. 2013; 8(1125): 3-21. (In Russian).
56. IDF Diabetes Atlas 10th edition International Diabetes Federation. 2021.
57. Di Angelantonio E, Bhupathiraju ShN, Wormser D, Gao P, Kaptoge S, Berrington de Gonzalez A, Cairns B, Huxley R, Jackson C, Joshy G, Lewington S, Manson J, Murphy N, Patel A, Samet J, Woodward M, Zheng W, Zhou M, Bansal N, Barricarte A, Carter B, James Cerhan, Smith G, Fang X, Franco O, Green J, Halsey J, Hildebrand J, Jung K, Korda R, McLerran D, Moore S, O'Keeffe L, Paige E, Ramond A, Reeves G, Rolland B, Sacerdote C, Sattar N, Sofianopoulou E, Stevens J, Thun M, Ueshima H, Yang L, Yun Y, Willeit P, Banks E, Valerie Beral, Chen Z, Gapstur S, Gunter M, Hartge P, Jee S, Lam TH, Peto R, Potter J, Willett W, Thompson S, Danesh J, Hu F. Body-mass index and all-cause mortality: individual-participant-data meta-analysis of 239 prospective studies in four continents. *Lancet*. 2016; 388(10046): 776-786.
58. Saienko YaA, Zak KP, Popova VV, Semionova TA. Leukocyte composition and immunophenotype of the blood lymphocytes in women with type 2 diabetes mellitus and obesity. *Int J Endocrinol*. 2016; (5(77)): 13-19. (In Russian).
59. Arbel Y, Finkelstein A, Halkin A, Birati EY, Revivo M, Zuzut M, Shevach A, Berliner S, Herz I, Keren G, Banai S. Neutrophil/lymphocyte ratio is related to the severity of coronary artery disease and clinical outcome in patients undergoing angiography. *Atherosclerosis*. 2012; 225(2): 456-460.
60. Wang Y, Xiao Y, Zhong L, Ye D, Zhang J, Tu Y, Bornstein SR, Zhou Z, Lam KSL, Xu A. Increased neutrophil elastase and proteinase 3 and augmented NETosis are closely associated with β -cell autoimmunity in patients with type 1 diabetes. *Diabetes*. 2014; 63(12): 4239-4248.
61. Leslie RD, Palmer J, Schloot NC, Lernmark A. Diabetes at the crossroads: relevance of disease classification to pathophysiology and treatment. *Diabetologia*. 2016; 59(1): 13-20.
62. Silvestri M, Bontempelli M, Giacomelli M, Malerba M, Rossi GA, Di Stefano A, Rossi A, Ricciardolo FLM. High serum levels of tumour necrosis factor-alpha and interleukin-8 in severe asthma: markers of systemic inflammation? *Clin Exp Allergy*. 2006; 36(11): 1373-1381.
63. Ponti G, Maccaferri M, Ruini C, Tomasi A, Ozben T. Biomarkers associated with COVID-19 disease progression. *Crit Rev Clin Lab Sci*. 2020; 57(6): 389-399.
64. Bouter KP, Meyling FH, Hoekstra JB, Masurel N, Erkelens DW, Diepersloot RJ. Influence of blood glucose levels on peripheral lymphocytes in patients with diabetes mellitus. *Diabetes Res*. 1992; 19(2): 77-80.
65. Dalmas E, Venteclef N, Caer C, Poitou C, Cremer I, Aron-Wisnewsky J, Lacroix-Desmazes S, Bayry J, Kaveri SV, Clément K, André S, Guerre-Millo M. T cell-derived IL-22 amplifies IL-1 β -driven inflammation in human adipose tissue: relevance to obesity and type 2 diabetes. *Diabetes*. 2014; 63(6): 1966-1977.
66. O'Rourke RW, Kay T, Lyle EA, Traxler SA, Deveney CW, Jobe BA, Roberts CT Jr, Marks D, Rosenbaum JT. Alterations in peripheral blood lymphocyte cytokine expression in obesity. *Clin Exp Immunol*. 2006; 146(1): 39-46.
67. Womack J, Tien PC, Feldman J, Shin JH, Fennie K, Anastos K, Cohen MH, Bacon MC, Minkoff H. Obesity and immune cell counts in women. *Metabolism*. 2007; 56(7): 998-1004.
68. Che TT, Ren Y, Liu SF. Expression of circulating CD4⁺ CD25⁺ FOXP3⁺ regulation T cells in obese patients. *Diabetologia*. 2013; 56(Suppl 1): A563.
69. Roitt IM, Brostoff J, Male D. Immunology.(5th edition). Mosby, London-Tokyo, 1998. 581 p.
70. Afanasieva VV, Zak KP, Butenko AK. Electron microscopy and ultracytochemistry of blood lymphocytes containing Gall bodies in healthy individuals. *Tsitol Genet*. 2003; 37(1): 56-60. (In Russian).
71. Velardi A, Grossi CE, Cooper MD. A large subpopulation of lymphocytes with T helper phenotype (Leu-3/T4⁺) exhibits the property

- of binding to NK cell targets and granular lymphocyte morphology. *J Immunol.* 1985; 134(1): 58-64.
72. Khomenko BM, Gruzov MA, Shlyakhovenko VS, Zak KP. The content and ultrastructure of blood CD4⁺ lymphocytes in healthy people and patients with type 1 diabetes. *Fiziol Zhurn.* 1989; 35(5): 31-38.
 73. Vozianov AF, Butenko AK, Zak KP. Cytokines. Biological and antitumor properties. Kiev: Naukova Dumka, 1998. 315 p.
 74. Wang Y, van Boxel-Dezaire AH, Cheon H, Yang J, Stark GR. STAT3 activation in response to IL-6 is prolonged by the binding of IL-6 receptor to EGF receptor. *Proc Natl Acad Sci USA.* 2013; 110(42): 16975-16980.
 75. Dror E, Dalmas E, Meier DT, Wuest S, Thévenet J, Thienel C, Timper K, Nordmann TM, Traub S, Schulze F, Item F, Vallois D, Pattou F, Kerr-Conte J, Lavallard V, Berney T, Thorens B, Konrad D, Böni-Schnetzler M, Donath MY. Postprandial macrophage-derived IL-1 β stimulates insulin, and both synergistically promote glucose disposal and inflammation. *Nat Immunol.* 2017; 18(3): 283-292.
 76. Donath MY. Targeting inflammation in the treatment of type 2 diabetes: time to start. *Nat Rev Drug Discov.* 2014; 13(6): 465-476.
 77. Chae JS, Paik JK, Kang R, Kim M, Choi Y, Lee SH, Lee JH. Mild weight loss reduces inflammatory cytokines, leukocyte count, and oxidative stress in overweight and moderately obese participants treated for 3 years with dietary modification. *Nutr Res.* 2013; 33(3): 195-203.
 78. Herder C, Brunner E, Tabak A. Elevated levels of the anti-inflammatory interleukin-1 receptor antagonist (IL-1Ra) precede, but do not prevent, the onset of type 2 diabetes (The Whitehall II Study). *Diabetologia.* 2008; 51(Suppl 1): 313.
 79. Carstensen M, Herder C, Kivimäki M, et al. Acceleration of the interleukin-1 receptor antagonist (IL-1Ra) trajectory precedes the diagnosis of type 2 diabetes by 6 years: the Whitehall II prospective cohort study. *Diabetologia.* 2009; 52(Suppl 1): 22.
 80. Urwyler SA, Schuetz P, Ebrahimi F, Donath MY, Christ-Crain M. Interleukin-1 Antagonism Decreases Cortisol Levels in Obese Individuals. *J Clin Endocrinol Metab.* 2017; 102(5): 1712-1718.
 81. Thorand B, Kolb H, Baumert J, Koenig W, Chambless L, Meisinger C, Illig T, Martin S, Herder C. Elevated levels of interleukin-18 predict the development of type 2 diabetes: results from the MONICA/KORA Augsburg Study, 1984-2002. *Diabetes.* 2005; 54(10): 2932-2938.
 82. Daniele G, Guardado Mendoza R, Winnier D, Fiorentino TV, Pengou Z, Cornell J, Andreozzi F, Jenkinson C, Cersosimo E, Federici M, Tripathy D, Folli F. The inflammatory status score including IL-6, TNF- α , osteopontin, fractalkine, MCP-1 and adiponectin underlies whole-body insulin resistance and hyperglycemia in type 2 diabetes mellitus. *Acta Diabetol.* 2014; 51(1): 123-131.
 83. Rodrigues KF, Pietrani NT, Bosco AA, Campos FMF, Sandrim VC, Gomes KB. IL-6, TNF- α , and IL-10 levels/polymorphisms and their association with type 2 diabetes mellitus and obesity in Brazilian individuals. *Arch Endocrinol Metab.* 2017; 61(5): 438-446.
 84. Goyal R, Faizy AF, Siddiqui SS, Singhai M. Evaluation of TNF- α and IL-6 Levels in Obese and Non-obese Diabetics: Pre- and Postinsulin Effects. *N Am J Med Sci.* 2012; 4(4): 180-184.
 85. Roohi A, Tabrizi M, Abbasi F, Ataie-Jafari A, Nikbin B, Larijani B, Qorbani M, Meysamie A, Asgarian-Omran H, Nikmanesh B, Bajouri A, Shafiey N, Maleki A. Serum IL-17, IL-23, and TGF- β levels in type 1 and type 2 diabetic patients and age-matched healthy controls. *Biomed Res Int.* 2014; 2014: 718946.
 86. Chen C, Shao Y, Wu X, Huang C, Lu W. Elevated interleukin-17 levels in patients with newly diagnosed type 2 diabetes mellitus. *Biochem Physiol.* 2016; 5(2): 206.
 87. Mishima Y, Kuyama A, Tada A, Takahashi K, Ishioka T, Kibata M. Relationship between serum tumor necrosis factor- α and insulin resistance in obese men with Type 2 diabetes mellitus. *Diabetes Res Clin Pract.* 2001; 52(2): 119-123.
 88. Mirza S, Hossain M, Mathews C, Martinez P, Pino P, Gay JL, Rentfro A, McCormick JB, Fisher-Hoch SP. Type 2-diabetes is associated with elevated levels of TNF- α , IL-6 and adiponectin and low levels of leptin in a population of Mexican Americans: a cross-sectional study. *Cytokine.* 2012; 57(1): 136-142.

89. Hotamisligil GS. The role of TNF α and TNF receptors in obesity and insulin resistance. *J Intern Med.* 1999; 245(6): 621-625.
90. Seyhan A, Nunes-Lopez Yu, Garufi G. Differences in serum cytokine concentration in lean and obese individuals with prediabetes and type 2 diabetes. *Diabetes.* 2015; 64(Suppl 1): A472, 1825-P.
91. Williams A, Radford J, O'Brien J, Davison K. Type 2 diabetes and the medicine of exercise: The role of general practice in ensuring exercise is part of every patient's plan. *Aust J Gen Pract.* 2020; 49(4): 189-193.
92. De Graaf DM, Teufel LU, Joosten LAB, Dinarello CA. Interleukin-38 in Health and Disease. *Cytokine.* 2022; 152: 155824.
93. Ho KT, Shiau MY, Chang YH, Chen CM, Yang SC, Huang CN. Association of interleukin-4 promoter polymorphisms in Taiwanese patients with type 2 diabetes mellitus. *Metabolism.* 2010; 59(12): 1717-1722.
94. Silva-Filho JL, Caruso-Neves C, Pinheiro AAS. IL-4: an important cytokine in determining the fate of T cells. *Biophys Rev.* 2014; 6(1): 111-118.
95. Badr E, Assar M, Elshayeb EI, Fath El-Bab S, El-Kousy S. A preliminary study of the relation between IL-4 and hypertension in type II diabetes mellitus. *Mol Biol Rep.* 2018; 45(6): 1967-1972.
96. Sartangelo C, Marrchetti P, Marselli L. Suppressors of cytokine signaling (SOCS) in cytokine-induced human islet cell damage. Abstracts of the 37th Annual Meeting of the EASD Glasgow, United Kingdom, 9–13 September 2001. *Diabetologia.* 2001; 44(Suppl 1): A1-A325.
97. te Velde AA, Huijbens RJ, Heije K, de Vries JE, Figdor CG. Interleukin-4 (IL-4) inhibits secretion of IL-1 beta, tumor necrosis factor alpha, and IL-6 by human monocytes. *Blood.* 1990; 76(7): 1392-1397.
98. Cheung DL, Hart PH, Vitti GF, Whitty GA, Hamilton JA. Contrasting effects of interferon-gamma and interleukin-4 on the interleukin-6 activity of stimulated human monocytes. *Immunology.* 1990; 71(1): 70-75.
99. Binisor ID, Moldovan R, Moldovan I, Andrei AM, Banita MI. Abdominal Obesity and Type 2 Diabetes Mellitus are Associated With Higher Seric Levels of IL 4 in Adults. *Curr Health Sci J.* 2016; 42(3): 231-237.
100. Shiau MY, Chuang PH, Yang CP, Hsiao CW, Chang SW, Chang KY, Liu TM, Chen HW, Chuang CC, Yuan SY, Chang YH. Mechanism of Interleukin-4 Reducing Lipid Deposit by Regulating Hormone-Sensitive Lipase. *Sci Rep.* 2019; 9(1): 11974.
101. Alsaïd A, El-Missiry M, Hatata el-S, Tarabay M, Settin A. Association of IL-4-590 C>T and IL-13-1112 C>T gene polymorphisms with the susceptibility to type 2 diabetes mellitus. *Dis Markers.* 2013; 35(4): 243-247.
102. Iyer SS, Cheng G. Role of interleukin 10 transcriptional regulation in inflammation and autoimmune disease. *Crit Rev Immunol.* 2012; 32(1): 23-63.
103. van Exel E, Gussekloo J, de Craen AJ, Frölich M, Bootsma-Van Der Wiel A, Westendorp RG, Leiden 85 Plus Study. Low production capacity of interleukin-10 associates with the metabolic syndrome and type 2 diabetes : the Leiden 85-Plus Study. *Diabetes.* 2002; 51(4): 1088-1092.
104. Esposito K, Nappo F, Giugliano F, Di Palo C, Ciotola M, Barbieri M, Paolisso G, Giugliano D. Meal modulation of circulating interleukin 18 and adiponectin concentrations in healthy subjects and in patients with type 2 diabetes mellitus. *Am J Clin Nutr.* 2003; 78(6): 1135-1140.
105. Canecki-Varžić S, Prpić-Križevac I, Mihaljević S, Bilić-Ćurčić I, Alkhamis T, Wagner J, Škrlec I, Barbić J. Association Between Interleukin-10 Gene (-1082g/A) Polymorphism and Type 2 Diabetes, Diabetes-Related Traits, and Microvascular Complications in the Croatian Population. *Acta Clin Croat.* 2018; 57(1): 71-81.
106. Alexandraki K, Piperi C, Kalofoutis C, Singh J, Alaveras A, Kalofoutis A. Inflammatory process in type 2 diabetes: The role of cytokines. *Ann NY Acad Sci.* 2006; 1084: 89-117.
107. Herder C, Baumert J, Thorand B, Martin S, Löwel H, Kolb H, Koenig W. Chemokines and incident coronary heart disease: results from the MONICA/KORA Augsburg case-cohort study, 1984-2002. *Arterioscler Thromb Vasc Biol.* 2006; 26(9): 2147-2152.
108. Zhang Z, Yuan W, Sun L, Szeto FL, Wong KE, Li X, Kong J, Li YC. 1,25-Dihydroxyvitamin D₃ targeting of NF-kappaB suppresses high glucose-induced MCP-1 expression in

- mesangial cells. *Kidney Int.* 2007; 72(2): 193-201.
109. Müller S, Martin S, Koenig W, Hanifi-Moghaddam P, Rathmann W, Haastert B, Giani G, Illig T, Thorand B, Kolb H. Impaired glucose tolerance is associated with increased serum concentrations of interleukin 6 and co-regulated acute-phase proteins but not TNF-alpha or its receptors. *Diabetologia.* 2002; 45(6): 805-812.
 110. Zozulińska D, Majchrzak A, Sobieska M, Wiktorowicz K, Wierusz-Wysocka B. Serum interleukin-8 level is increased in diabetic patients. *Diabetologia.* 1999; 42(1): 117-118.
 111. Shah R, Hinkle CC, Ferguson JF, Mehta NN, Li M, Qu L, Qu L, Lu Y, Putt ME, Ahima RS, Reilly MP. Fractalkine is a novel human adipochemokine associated with type 2 diabetes. *Diabetes.* 2011; 60(5): 1512-1518.
 112. Smieszek SP, Polymeropoulos VM, Polymeropoulos CM, Przychodzen BP, Birznieks G, Polymeropoulos MH. Elevated plasma levels of CXCL16 in severe COVID-19 patients. *Cytokine.* 2022; 152: 155810.
 113. Neurath MF. COVID-19 and immunomodulation in IBD. *Gut.* 2020; 69(7): 1335-1342.
 114. Komisarenko SV. Scientists' pursuit for SARS-CoV-2 coronavirus: strategies against pandemic. *Ukr Biochem J.* 2020; 92(6): 5-52.
 115. Tsymbalyuk VI, Tronko MD, Popova VV. Modern views on the pathogenetic aspects and treatment strategies of patients with diabetes mellitus and COVID-19. *Endocrinologia.* 2020; 25(Spec Is 2): 3-44.
 116. Hu B, Huang S, Yin L. The cytokine storm and COVID-19. *J Med Virol.* 2021; 93(1): 250-256.
 117. Bergantini L, d'Alessandro M, Cameli P, Otranto A, Luzzi S, Bianchi F, Bargagli E. Cytokine profiles in the detection of severe lung involvement in hospitalized patients with COVID-19: The IL-8/IL-32 axis. *Cytokine.* 2022; 151: 155804.
 118. Ashrafzadeh-Kian S, Campbell MR, Jara Aguirre JC, Walsh J, Kumanovics A, Jenkinson G, Rinaldo P, Snyder MR, Algeciras-Schimnich A. Role of immune mediators in predicting hospitalization of SARS-CoV-2 positive patients. *Cytokine.* 2022; 150: 155790.
 119. Shoenfeld Y. Corona (COVID-19) time musings: Our involvement in COVID-19 pathogenesis, diagnosis, treatment and vaccine planning. *Autoimmun Rev.* 2020; 19(6): 102538.
 120. Liu R, Wang Y, Li J, Han H, Xia Z, Liu F, Wu K, Yang L, Liu X, Zhu C. Decreased T cell populations contribute to the increased severity of COVID-19. *Clin Chim Acta.* 2020; 508: 110-114.
 121. Feldman EL, Savelieff MG, Hayek SS, Pennathur S, Kretzler M, Pop-Busui R. COVID-19 and Diabetes: A Collision and Collusion of Two Diseases. *Diabetes.* 2020; 69(12): 2549-2565.
 122. Metwally AA, Mehta P, Johnson BS, Nagarjuna A, Snyder MP. COVID-19-Induced New-Onset Diabetes: Trends and Technologies. *Diabetes.* 2021; 70(12): 2733-2744.
 123. Paul WE. Ed. Fundamental Immunology. 4th ed. Philadelphia: Lippincott-Raven, 1998. 1589 p.
 124. Li X, Geng M, Peng Y, Meng L, Lu S. Molecular immune pathogenesis and diagnosis of COVID-19. *J Pharm Anal.* 2020; 10(2): 102-108.
 125. Qun S, Tang Y, Sun J, Liu Z, Wu J, Zhang J, Guo J, Xu Z, Zhang D, Chen Z, Hu F, Xu X, Ge W. Neutrophil-To-Lymphocyte Ratio Predicts 3-Month Outcome of Acute Ischemic Stroke. *Neurotox Res.* 2017; 31(3): 444-452.
 126. Armstrong EJ, Morrow DA, Sabatine MS. Inflammatory biomarkers in acute coronary syndromes: part I: introduction and cytokines. *Circulation.* 2006; 113(6): e72-e75.
 127. Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, Si HR, Zhu Y, Li B, Huang CL, Chen HD, Chen J, Luo Y, Guo H, Jiang RD, Liu MQ, Chen Y, Shen XR, Wang X, Zheng XS, Zhao K, Chen QJ, Deng F, Liu LL, Yan B, Zhan FX, Wang YY, Xiao GF, Shi ZL. Addendum: A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature.* 2020; 588(7836): E6.
 128. Declercq J, De Leeuw E, Lambrecht BN. Inflammasomes and IL-1 family cytokines in SARS-CoV-2 infection: from prognostic marker to therapeutic agent. *Cytokine.* 2022; 157: 155934.
 129. Bramante CT, Huling JD, Tignanelli CJ, Buse JB, Liebovitz DM, Nicklas JM, Cohen K, Puskaric MA, Belani HK, Proper JL, Siegel LK, Klatt NR, Odde DJ, Luke DG, Anderson B, Karger AB, Ingraham NE, Hartman KM, Rao V, Hagen AA, Patel B, Fenno SL, Avula N, Reddy NV, Erickson SM, Lindberg S, Friction R, Lee S, Zaman A,

- Saveraid HG, Tordsen WJ, Pullen MF, Biros M, Sherwood NE, Thompson JL, Boulware DR, Murray TA. Randomized Trial of Metformin, Ivermectin, and Fluvoxamine for Covid-19. *N Engl J Med*. 2022; 387(7): 599-610.
1302. Todorović-Raković N, Whitfield JR. Between immunomodulation and immunotolerance: The role of IFN γ in SARS-CoV-2 disease. *Cytokine*. 2021; 146: 155637.
 131. Khunti K, Del Prato S, Mathieu C, Kahn SE, Gabbay RA, Buse JB. COVID-19, Hyperglycemia, and New-Onset Diabetes. *Diabetes Care*. 2021; 44(12): 2645-2655.
 132. Li J, Huang DQ, Zou B, Yang H, Hui WZ, Rui F, Yee NTS, Liu C, Nerurkar SN, Kai JCY, Teng MLP, Li X, Zeng H, Borghi JA, Henry L, Cheung R, Nguyen MH. Epidemiology of COVID-19: A systematic review and meta-analysis of clinical characteristics, risk factors, and outcomes. *J Med Virol*. 2021; 93(3): 1449-1458.
 133. Atkinson MA, Powers AC. Distinguishing the real from the hyperglycaemia: does COVID-19 induce diabetes? *Lancet Diabetes Endocrinol*. 2021; 9(6): 328-329.
 134. Müller JA, Groß R, Conzelmann C, Krüger J, Merle U, Steinhart J, Weil T, Koepke L, Bozzo CP, Read C, Fois G, Eiseler T, Gehrmann J, van Vuuren J, Wessbecher IM, Frick M, Costa IG, Breunig M, Grüner B, Peters L, Schuster M, Liebau S, Seufferlein T, Stenger S, Stenzinger A, MacDonald PE, Kirchhoff F, Sparrer KMJ, Walther P, Lickert H, Barth TFE, Wagner M, Münch J, Heller S, Kleger A. SARS-CoV-2 infects and replicates in cells of the human endocrine and exocrine pancreas. *Nat Metab*. 2021; 3(2): 149-165.
 135. Tang X, Uhl S, Zhang T, Xue D, Li B, Vandana JJ, Acklin JA, Bonnycastle LL, Narisu N, Erdos MR, Bram Y, Chandar V, Chong ACN, Lacko LA, Mi Z, Lim JK, Borczuk AC, Xiang J, Naji A, Collins FS, Evans T, Liu C, tenOever BR, Schwartz RE, Chen S. SARS-CoV-2 infection induces beta cell transdifferentiation. *Cell Metab*. 2021; 33(8): 1577-1591.e7.
 136. Rathmann W, Kuss O, Kostev K. Incidence of newly diagnosed diabetes after Covid-19. *Diabetologia*. 2022; 65(6): 949-954.
 137. Crouse AB, Grimes T, Li P, Might M, Ovalle F, Shalev A. Metformin Use Is Associated With Reduced Mortality in a Diverse Population With COVID-19 and Diabetes. *Front Endocrinol (Lausanne)*. 2021; 11: 600439.
 138. Varghese E, Samuel SM, Liskova A, Kubatka P, Büsselberg D. Diabetes and coronavirus (SARS-CoV-2): Molecular mechanism of Metformin intervention and the scientific basis of drug repurposing. *PLoS Pathog*. 2021; 17(6): e1009634.
 139. Zak KP, Furmanova OV, Popova VV, Sayenko YaA. The content of pro-inflammatory cytokines IL-1 β , IL-6, IL-17A and TNF α in the blood of patients with type 2 diabetes after therapy with metformin. *Ukr Biochem J*. 2020; 92(6): 105-112.
 140. Horby P, Mafham M, Linsell L, Bell JL, Staplin N, Emberson JR, Wiselka M, Ustianowski A, Elmahi E, Prudon B, Whitehouse T, Felton T, Williams J, Faccenda J, Underwood J, Baillie JK, Chappell LC, Faust SN, Jaki T, Jeffery K, Lim WS, Montgomery A, Rowan K, Tarning J, Watson JA, White NJ, Juszczak E, Haynes R, Landray MJ. Effect of Hydroxychloroquine in Hospitalized Patients with Covid-19. *N Engl J Med*. 2020; 383(21): 2030-2040.
 141. Horby P, Lim WS, Emberson JR, Mafham M, Bell JL, Linsell L, Staplin N, Brightling C, Ustianowski A, Elmahi E, Prudon B, Green C, Felton T, Chadwick D, Rege K, Fegan C, Chappell LC, Faust SN, Jaki T, Jeffery K, Montgomery A, Rowan K, Juszczak E, Baillie JK, Haynes R, Landray MJ. Dexamethasone in Hospitalized Patients with Covid-19. *N Engl J Med*. 2021; 384(8): 693-704.