

FIBROBLAST GROWTH FACTOR 23, CALCIUM AND PHOSPHATE SERUM LEVELS IN EXPERIMENTAL PREECLAMPSIA: IMPACT OF VITAMIN D₃ STATUS

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Received: 25 August 2025; **Revised:** 02 October 2025; **Accepted:** 30 January 2026

Preeclampsia (PE) is a major cause of maternal and perinatal morbidity, with its pathogenesis involving, in particular, impaired mineral homeostasis. Recent evidence suggest that fibroblast growth factor 23 (FGF23), a key regulator of phosphate balance and vitamin D₃ metabolism, may contribute to pregnancy complications, however, its role in PE remains poorly understood. This study aimed to evaluate serum FGF23 level and its relationship with vitamin D₃ and calcium–phosphate balance at preeclampsia development in animal experimental model. Thirty-five Wistar female rats were divided into three groups: controls on a standard diet; vitamin D₃-deficient rats; vitamin D₃-deficient rats supplemented with cholecalciferol. Within each group, PE was induced by N ω -nitro-L-arginine methyl ester administration. Serum concentrations of 25(OH)D₃, FGF23, parathyroid hormone (PTH), total calcium, inorganic phosphate, and alkaline phosphatase (ALP) activity were determined by ELISA and biochemical assays. It was shown that vitamin D₃ deficiency was accompanied by hypocalcemia, hypophosphatemia, elevated FGF23 and increased ALP activity in the serum. Supplementation with vitamin D₃ increased 25(OH)D₃, markedly reduced FGF23 levels and normalized mineral parameters. Induction of PE caused significant disturbances in calcium–phosphate status, hypertension, and 100% mortality of vitamin D₃-deficient animals. In the presence of preeclampsia vitamin D₃ efficacy was limited. In PE group, despite of vitamin D₃ supplementation, serum FGF23 was markedly elevated, indicating impaired vitamin D₃ metabolism. Our findings demonstrate that vitamin D₃ deficiency amplifies PE severity through disruption of mineral homeostasis and FGF23 dependent signaling.

Key words: 25(OH)D₃, vitamin D₃ deficiency, L-NAME, preeclampsia, calcium–phosphate metabolism.

Preeclampsia (PE) remains one of the leading causes of maternal and perinatal morbidity and mortality worldwide, representing a major medical and social challenge in modern obstetrics [1, 2]. This multifactorial syndrome, typically manifesting after the 20th week of gestation, is characterized by newly diagnosed arterial hypertension in combination with proteinuria and is accompanied by a wide spectrum of systemic disorders. The pathogenesis of preeclampsia involves complex interactions among defective vascular remodeling, endothelial dysfunction, systemic inflammation, and immune response imbalance [3, 4]. Despite substantial progress in understanding risk factors and

molecular mechanisms underlying this condition, highly specific and sensitive biomarkers for its early detection and prediction, as well as effective preventive strategies with proven clinical benefit, are still lacking.

In recent years, there has been growing interest in the role of hormonal regulators of calcium–phosphate metabolism in the pathogenesis of pregnancy complications, particularly preeclampsia [5, 6]. Of special interest is fibroblast growth factor 23 (FGF23), a hormone-like protein predominantly secreted by osteocytes and osteoblasts, which plays a key role in maintaining phosphate homeostasis and regulating vitamin D₃ metabolism [7-9]. At the level

of renal proximal tubules, FGF23 reduces phosphate reabsorption by suppressing the sodium–phosphate co-transporters NaPi-IIa and NaPi-IIc, while simultaneously inhibiting the activity of 1α -hydroxylase 25-hydroxyvitamin D (CYP27B1) – the enzyme responsible for the synthesis of the biologically active form of vitamin D₃ (calcitriol, $1\alpha,25(\text{OH})_2\text{D}_3$). In parallel, it induces the expression of 24-hydroxylase (CYP24A1), which catalyzes the degradation of vitamin D₃, thereby decreasing its bioavailability. These effects are mediated through the activation of intracellular signaling cascades (mainly ERK1/2) following the interaction of FGF23 with its receptor FGFR1 in the presence of the co-receptor α -Klotho.

Experimental and clinical evidence indicates that excessive FGF23 activity leads to a significant reduction in $1\alpha,25(\text{OH})_2\text{D}_3$ concentrations, both through the inhibition of its synthesis and through enhanced catabolic degradation [10, 11]. Importantly, the impact of FGF23 is not confined to renal tissue but also extends to extra-renal sources of active vitamin D₃ production, which is of fundamental significance under conditions associated with its deficiency.

Vitamin D₃, represented both by its circulating form 25-hydroxyvitamin D₃ ($25(\text{OH})\text{D}_3$) and its biologically active metabolite $1\alpha,25(\text{OH})_2\text{D}_3$, performs a range of critically important functions during pregnancy: it regulates immunological tolerance, supports endothelial function, promotes placental vascularization, and facilitates trophoblast differentiation [12–15]. Maternal deficiency of $25(\text{OH})\text{D}_3$ has been associated with an increased risk of preeclampsia, intrauterine growth restriction, and preterm birth. Although meta-analyses and systematic reviews of clinical and experimental studies indicate a potential protective effect of adequate vitamin D₃ bioavailability, the results of interventional trials remain inconsistent due to variability in supplementation dosage, timing of administration, and population characteristics [16–18].

Data regarding alterations in FGF23 levels during pregnancy and in preeclampsia remain inconsistent. Some studies report its serological elevation in affected mothers, while others demonstrate a decrease [19]. These discrepancies may be attributed to differences in preeclampsia phenotypes (early- vs. late-onset and severity), timing of sample collection, and methodological aspects of measurement. Recent clinical studies show correlations between FGF23 concentrations and biomarkers of placental

dysfunction, suggesting its potential involvement in the pathogenesis of preeclampsia, although a comprehensive understanding of this association has yet to be fully established [20]. Experimental models indicate a possible influence of FGF23 on the expression of placental genes involved in vitamin D₃ metabolism (particularly Cyp24a1), as well as its capacity to indirectly modulate angiogenesis. At present, it remains relevant to elucidate the relationship between FGF23-mediated signaling, vitamin D₃ metabolism, and mineral homeostasis in both normal and pathological pregnancy.

A relevant scientific challenge is to elucidate the role of fibroblast growth factor 23 in the regulation of vitamin D₃ metabolism and its impact on pregnancy under both physiological and complicated conditions. In particular, the relationship between changes in FGF23 levels, disturbances in calcium–phosphate homeostasis, and the development of PE remains insufficiently understood. In this context, the objective of our study was to assess FGF23 levels during pregnancy and determine their association with preeclampsia in an experimental model.

Materials and Methods

Animals and general experimental design. The experimental cohort consisted of 35 eight-week-old female Wistar rats with a mean body weight of 174 ± 12 g. All animals were housed under standard vivarium conditions (temperature $22 \pm 2^\circ\text{C}$, relative humidity 55–60%, 12-h light/dark cycle) with unrestricted access to food and water.

The animals were allocated into three experimental groups (Fig. 1). Group I ($n = 10$) – control females fed a standard vivarium diet without vitamin D₃ restrictions. Group II ($n = 12$) – females fed a rachitogenic diet (wheat flour – 89.5%, CaCO₃ – 3%, NaCl – 2%, dried brewer's yeast – 0.01%, vitamin A – 0.01%, vitamin E – 0.01%, water-soluble vitamins (Complevit) – 0.02%) completely lacking vitamin D₃ for 60 days before mating to establish a vitamin D₃-deficient status. Group III ($n = 13$) – females subjected to the same vitamin D₃-deficient diet for 60 days, followed by transition to a regular laboratory diet supplemented with oral vitamin D₃. Cholecalciferol (Sigma, USA) was administered in sunflower oil via gavage at a dose of 1000 IU/kg body weight (0.2 ml oil solution) daily for two weeks prior to mating.

Ethical statement. All experimental procedures were conducted following established bioethi-



Fig. 1. Experimental timeline for rat groups

cal principles and international regulations for the use of laboratory animals, including EU Directive 2010/63/EU, the European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes (Strasbourg, 1986), and the Ukrainian Guidelines for Bioethical Assessment of Preclinical and Other Scientific Research on Animals (Kyiv, 2006). The study was approved by the Bioethics Committee of Bogomolets National Medical University (Protocol No. 123, November 4, 2024).

Induction of vitamin D₃ deficiency. Groups II and III were fed a vitamin D₃-deficient rachitogenic diet for 60 days prior to mating to induce deficiency [16]. Blood samples were collected from the tail vein to assess 25-hydroxyvitamin D₃ (25(OH)D₃) levels using a General 25-Hydroxyvitamin D₃ ELISA kit (UNDL00047, Assay Genie, Ireland), and concentrations were calculated with Gain Data® (Arigo's ELISA Calculator).

Mating and pregnancy monitoring. After confirming vitamin D₃ deficiency, the females were mated with fertile males at a male-to-female ratio of 2:1. The presence of spermatozoa in vaginal smears the next morning was used to define gestational day 1 (GD1).

Induction of preeclampsia. To induce L-NAME-mediated preeclampsia (PE), each experimental group was divided into subgroups A and B. Animals in subgroup B received subcutaneous injections of L-NAME (N ω -nitro-L-arginine methyl ester) at a dose of 100 mg/kg body weight on GD5 and GD7 [21]. From GD5 to GD15, the animals were further administered L-NAME daily in drinking water at a dose of 40 mg/kg body weight. Rats in subgroup A continued on the respective diet and received two subcutaneous injections of physiological saline (vehicle).

Blood pressure measurement. Blood pressure was measured using the CODA® Monitor (Kent Scientific, USA), a non-invasive tail-cuff system based on volume pressure recording (VPR). Measurements were taken on GD1 (baseline) and GD15 (following 10 days of L-NAME treatment). Rats were placed in restrainers on a thermostatically controlled platform (37–38°C) to ensure stable blood flow, and 3–5 readings were obtained within 5–7 minutes to minimize variability.

Determination of calcium–phosphate metabolism and related biomarkers. The serum content of FGF23 in pregnant rats was measured using an enzyme-linked immunosorbent assay (ELISA) with the Rat FGF23 (Fibroblast Growth Factor 23) ELISA kit (RTFI00086; AssayGenie, Ireland) according to the manufacturer's protocol.

Parathyroid hormone (PTH) levels were determined by ELISA using the Rat Pth (Parathyroid Hormone) ELISA kit (RTFI00259; AssayGenie, Ireland) following the manufacturer's instructions. The concentrations of FGF23 and PTH in the analyzed samples were calculated using GainData® (arigo's ELISA Calculator).

Serum calcium levels were assessed with a commercial bio-test kit (Lachema, Czech Republic), using a standard solution of 25 mmol/l CaCO₃ dissolved in 1.7% HCl. The method is based on the ability of calcium to form, in an alkaline medium, a red-colored complex with glyoxal-bis-2(oxyanil), which was quantified spectrophotometrically at $\lambda = 540$ nm.

The concentration of inorganic phosphate (Pi) in serum was determined after protein precipitation with 12% trichloroacetic acid (TCA) using the Dyce method. Protein-free extracts (0.5 ml) were incubated at 37°C for 10 min with ascorbic acid (260 mM) and ammonium molybdate (7 mM). After incubation, the samples were placed on ice for 10 min, and optical density was measured spectrophotometrically at $\lambda = 640$ nm.

The activity of alkaline phosphatase (alkaline phosphomonoesterase, orthophosphoric acid monoester hydrolase) in serum was determined by measuring the formation of 4-nitrophenol resulting from the hydrolysis of the substrate 4-nitrophenylphosphate. The reaction was stopped by adding 30 mM Trilon B solution in 1 M NaOH. The optical density of the samples was measured spectrophotometrically at $\lambda = 410$ nm. The activity of total alkaline phosphatase was calculated using the following formula:

$$X = \varepsilon(\text{sample})/\varepsilon(\text{standard}) \times K,$$

where X is the number of micromoles of 4-nitrophenol released by the enzyme per liter of serum per minute at 37°C; 0.096 – mmol of 4-nitrophenylphosphate in 0.04 ml of standard solution; 0.04 – serum volume (ml); 15 – incubation time (min).

The activity of the bone-specific alkaline phosphatase isoform was determined as the difference between the total alkaline phosphatase activity and the thermostable alkaline phosphatase activity, after heating the samples in a water bath at 56–57°C.

Statistical analysis. Data were analyzed using conventional methods of variation statistics, with the calculation of the mean (M) and standard error of the mean ($\pm m$). Statistical significance of differences between groups was assessed using one-way ANOVA followed by Bonferroni post hoc correction. Differences were considered significant at $P < 0.05$. Statistical processing was performed using Origin-Lab 8.5 software.

Results and Discussion

Serum 25-hydroxyvitamin D₃, the primary hydroxylated metabolite of cholecalciferol, serves as a reliable marker of systemic vitamin D₃ bioavailability due to its prolonged half-life (2–3 weeks), reflecting both endogenous synthesis and dietary or supplemental intake. In the present study, we observed that

serum 25(OH)D₃ concentrations in pregnant female rats were significantly modulated by dietary regimen (standard versus rachitogenic), exogenous vitamin D₃ supplementation, and the presence of experimental preeclampsia, highlighting the combined influence of nutritional status and pathological conditions on vitamin D₃ homeostasis during gestation.

As shown in Fig. 2, in animals maintained on a vitamin D₃-deficient diet (Group IIA), the mean serum 25(OH)D₃ level was 16.5 ± 1.2 ng/ml, which was nearly threefold lower ($P < 0.001$) compared with the control Group IA (49.6 ± 2.4 ng/ml) and corresponds to the development of severe vitamin D₃ deficiency (Holick, 2011). In Group IIB (vitamin D₃ deficiency with preeclampsia), all animals died, indicating high mortality associated with the combination of profound vitamin D₃ deficiency and preeclampsia. Consequently, biochemical data for this group were not obtained.

Administration of vitamin D₃ to rats (Group IIIA) resulted in a significant increase in 25(OH)D₃ levels to 34.6 ± 2.1 ng/ml ($P < 0.01$ vs. IA); however, in Group IIIB (with preeclampsia), 25(OH)D₃ decreased to 30.7 ± 1.2 ng/ml ($P < 0.05$ vs. IIIA), indicating partial attenuation of vitamin D₃ supplementation efficacy in the presence of preeclampsia. Intact animals (Group IA) exhibited the highest serum 25(OH)D₃ levels (49.6 ± 2.4 ng/ml), whereas the

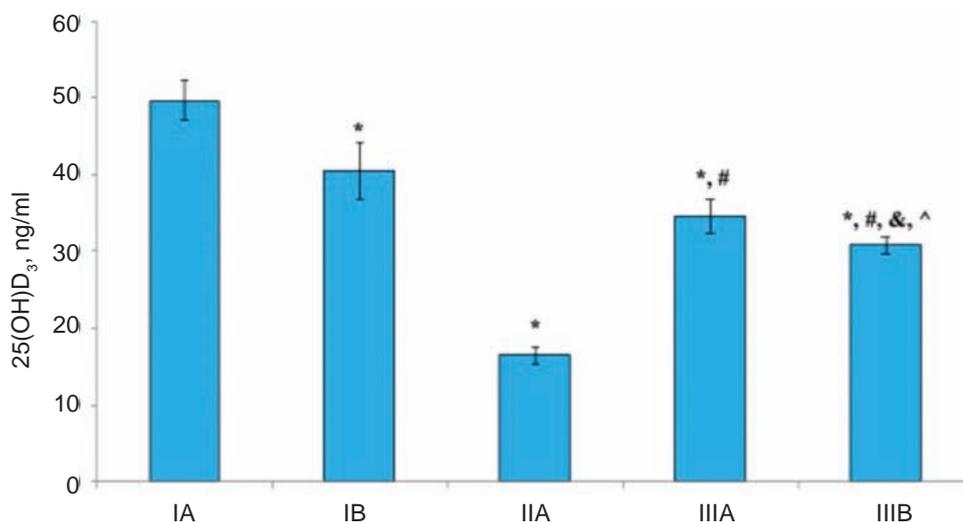


Fig. 2. Serum 25(OH)D₃ levels in the experimental groups ($M \pm m$): IA – control group without preeclampsia, IB – control group + preeclampsia, IIA – vitamin D₃-deficiency, without preeclampsia, IIIA – vitamin D₃-deficiency + vitamin D₃, without preeclampsia, IIIB – vitamin D₃-deficiency + vitamin D₃ + preeclampsia. *Statistically significant difference compared to Group IA; #statistically significant difference compared to Group IIA; &statistically significant difference compared to Group IIIA; ^statistically significant difference compared to Group IB, $P < 0.05$

development of preeclampsia in this group (Group IB) was associated with a reduction to 40.4 ± 3.6 ng/ml ($P < 0.01$ vs. control).

One of the key diagnostic criteria for preeclampsia in the studied groups was the presence of hemodynamic alterations, specifically hypertension, as presented in Fig. 3.

In the group of animals with vitamin D₃ deficiency (IIA), on gestational day 15 (GD15), mean systolic/diastolic blood pressure values were $132.00 \pm 2.31/88.90 \pm 2.04$ mmHg, which was significantly higher ($P < 0.01$) compared with control rats in Group IA ($110.20 \pm 4.56/69.60 \pm 4.01$ mmHg). In Group IIB (vitamin D₃ deficiency + preeclampsia), all animals died prior to blood pressure measurement, confirming the severe progression of preeclampsia induced by L-NAME in the context of vitamin D₃ deficiency.

Vitamin D₃ supplementation in Group IIIA (without preeclampsia) was associated with a reduction in blood pressure to $118.50 \pm 4.27/77.00 \pm 1.69$ mmHg ($P < 0.05$ vs. IIA), indicating a beneficial effect of vitamin D₃ correction on vascular tone and hemodynamics. However, in the presence of preeclampsia with vitamin D₃ supplementation (Group IIIB), blood pressure increased to $174.70 \pm 5.11/118.00 \pm 5.67$ mmHg ($P < 0.001$ vs. IIIA), which exceeded even the hypertension observed in Group IIA. In the control Group IA, blood pressure remained within the physiological range ($110.20 \pm 4.56/69.60 \pm 4.01$

mmHg), whereas the development of preeclampsia in Group IB was accompanied by an increase to $163.80 \pm 2.76/105.20 \pm 3.57$ mmHg ($P < 0.001$ vs. intact controls). These findings indicate that the combination of vitamin D₃ deficiency and preeclampsia leads to more pronounced hemodynamic disturbances than any other experimental condition studied in this model.

Considering the direct involvement of the hormonally active form of vitamin D₃ in the formation of the inorganic bone matrix and in the regulation of calcium–phosphate metabolism, we further analyzed changes in blood mineral components. The observed alterations were generally consistent with the vitamin D₃ status of pregnant rats under normal conditions, as well as during the induction of nutritional vitamin D₃ deficiency and L-NAME-induced preeclampsia (Table).

In Group IIA, total serum calcium was decreased to 1.87 ± 0.02 μmol/min-l compared with 2.30 ± 0.04 μmol/min-l in intact controls ($P < 0.001$), reflecting the development of classical hypocalcemia under conditions of low vitamin D₃ status. In Groups IIIA and IIIB, calcium levels increased to 1.92 ± 0.02 and 1.94 ± 0.02 μmol/min-l, respectively, correlating with elevated serum 25(OH)D₃ concentrations and indicating partial restoration of calcium homeostasis following vitamin D₃ supplementation. In control animals with induced preeclampsia (Group IB), serum calcium decreased significantly to 2.10 ± 0.01 μmol/

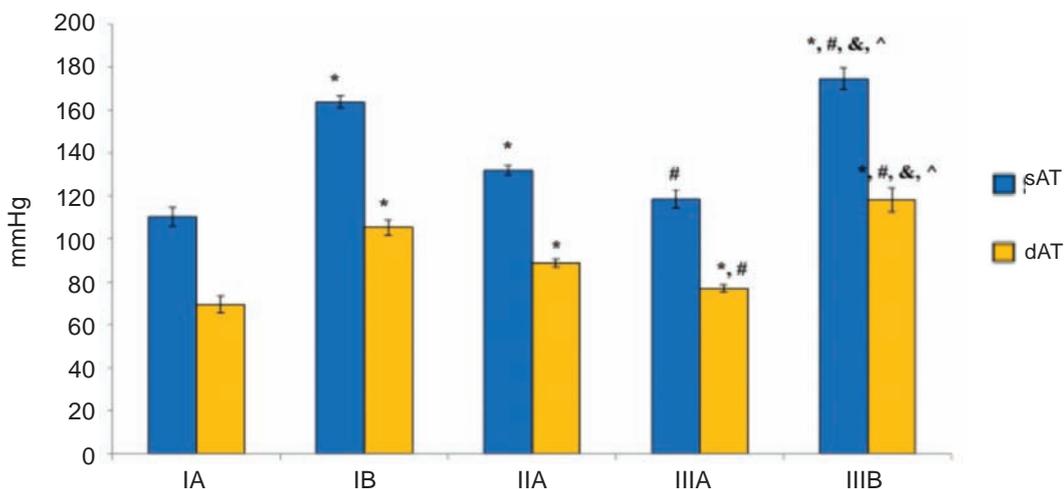


Fig. 3. Blood pressure characteristics in the experimental groups ($M \pm m$): IA – control group without preeclampsia, IB – control group + preeclampsia, IIA – vitamin D₃-deficiency, without preeclampsia, IIIA – vitamin D₃-deficiency + vitamin D₃, without preeclampsia, IIIB – vitamin D₃-deficiency + vitamin D₃ + preeclampsia. *Statistically significant difference vs. Group IA; #statistically significant difference vs. Group IIA; &statistically significant difference vs. Group IIIA; ^statistically significant difference vs. Group IB, $P < 0.05$

Table. Calcium–phosphate metabolism parameters

Indicator	Control (Group I), <i>n</i> = 10		D ₃ hypovitaminosis (Group II), <i>n</i> = 6	D ₃ hypovitaminosis + vitamin D ₃ (Group III), <i>n</i> = 13	
	IA (without PE), <i>n</i> = 5	IB (PE), <i>n</i> = 5	IIA (without PE), <i>n</i> = 6	IIIA (without PE), <i>n</i> = 7	IIIB (PE), <i>n</i> = 6
Total calcium, μmol/min·l	2.30 ± 0.04	2.10 ± 0.01 [^]	1.87 ± 0.02 [*]	1.92 ± 0.02 [#]	1.94 ± 0.02 ^{&}
Inorganic phosphate, μmol/min·l	2.15 ± 0.03	2.29 ± 0.02 [^]	1.80 ± 0.03 [*]	2.02 ± 0.07 [#]	2.15 ± 0.04 ^{&}
Total alkaline phosphatase, μmol/min·l	207.6 ± 1.5	276.90 ± 3.29 [^]	325.6 ± 2.4 [*]	244.40 ± 3.84 [#]	347.70 ± 7.01 ^{&}
Bone isoenzyme of alkaline phosphatase, μmol/min·l	196.30 ± 3.03	259.5 ± 5.6 [^]	302.80 ± 4.05 [*]	236.20 ± 3.97 [#]	327.40 ± 7.98 ^{&}

Note. Group IIB(PE) – all animals died demonstrating extremely high mortality associated with vitamin D₃ deficiency and preeclampsia. ^{*}Statistically significant difference vs. IA; [#]statistically significant difference vs. IIA; [&]statistically significant difference vs. IIIA; [^]statistically significant difference vs. IB; *P* < 0.05

min·l compared with rats not receiving L-NAME (*P* < 0.05).

Serum inorganic phosphate concentration in vitamin D₃-deficient animals (Group IIA) decreased to 1.80 ± 0.03 μmol/min·l compared with 2.15 ± 0.03 μmol/min·l in intact controls (*P* < 0.05; Table). Correction of vitamin D₃ deficiency with cholecalciferol in Group IIIA resulted in the Pi elevation to 2.02 ± 0.07 μmol/min·l, while in the context of preeclampsia induction (Group IIIB), levels rose to 2.15 ± 0.04 μmol/min·l (*P* < 0.05). In control animals, the development of preeclampsia (Group IB) was associated with a further increase in Pi to 2.29 ± 0.02 μmol/min·l (*P* < 0.05 vs. IA), which likely reflects the adverse effects of preeclampsia and renal dysfunction on phosphate excretion.

In the control group (I), the mean total alkaline phosphatase (ALP) activity was 207.6 ± 1.5 μmol/min·l, whereas in animals with vitamin D₃ deficiency (II), this parameter increased to 276.9 ± 3.29 μmol/min·l. The highest values were observed in the vitamin D₃-supplemented group (III) – 325.6 ± 2.40 μmol/min·l (*P* < 0.05 vs. control), which may reflect compensatory enhancement of mineralization processes in response to restored vitamin D status. Analysis of subgroups differing by preeclampsia status showed that in animals without PE (IA, IB, IIA, IIIA), total ALP activity ranged from 244.4

to 347.7 μmol/min·l, whereas in the presence of PE (IIIB) an additional increase to 347.7 ± 7.01 μmol/min·l was observed.

Bone-specific ALP isoenzyme demonstrated a similar pattern: 196.3 ± 3.03 μmol/min·l in controls, 259.5 ± 5.60 μmol/min·l in the vitamin D₃-deficient group, and 302.8 ± 4.05 μmol/min·l in the vitamin D₃-supplemented group (*P* < 0.05 vs. control). In the IIIB subgroup (PE), this parameter reached 327.4 ± 7.98 μmol/min·l, exceeding values in animals without PE, indicating a possible contribution of pathological pregnancy to bone tissue remodeling and osteoblast activation.

Increased activity of total alkaline phosphatase, particularly its bone isoform, may reflect disturbances in the deposition of calcium and phosphate in bone tissue and/or enhanced bone resorption as a key mechanism for mobilizing mineral components to maintain systemic calcium–phosphate homeostasis.

The classical response to hypocalcemia and hypophosphatemia associated with vitamin D₃ deficiency is the development of secondary hyperparathyroidism [22]. Analysis of serum parathyroid hormone levels demonstrated elevations above reference values for young non-pregnant female rats in all experimental groups. According to the literature, normal physiological PTH levels in non-pregnant females range from 15 to 65 pg/ml [23].

In vitamin D₃-deficient animals (Group IIA), PTH concentration reached 266.9 ± 8.7 pg/ml compared with 326.5 ± 13.0 pg/ml in the intact control group ($P < 0.05$; Fig. 4). Vitamin D₃ supplementation in Group IIIA resulted in an increase of PTH to 310.7 ± 9.8 pg/ml ($P < 0.05$ vs. IIA), likely reflecting activation of calcium metabolism and enhanced bone turnover. In Group IIIB, preeclampsia induction led to a decrease in PTH to 266.9 ± 15.6 pg/ml ($P < 0.05$ vs. IIIA), possibly highlighting significant renal impairment and suppression of PTH secretion.

In control rats (Group IA), PTH levels were the highest among all groups (326.5 ± 13.0 pg/ml), whereas preeclampsia induction (Group IB) reduced PTH to 272.7 ± 9.9 pg/ml ($P < 0.05$), confirming the probable inhibitory effect of L-NAME-induced preeclampsia on hormone secretion under the conditions of this experiment.

Fibroblast growth factor 23 (FGF23) is an important regulator of phosphate metabolism and placental function, with serum levels exhibiting significant and highly contrasting intergroup changes (Fig. 5).

The highest concentrations of this growth factor were observed in vitamin D₃-deficient animals (Group IIA), exceeding more than twice those of the intact control group (473.2 ± 10.59 vs. 208.2 ± 11.16 pg/ml, $P < 0.001$). This may reflect activation of the mechanisms for enhanced renal phosphate excretion and the development of phosphaturia under condi-

tions of vitamin D₃ deficiency. Partial correction of vitamin D₃ deficiency by cholecalciferol supplementation in Group IIIA elicited a marked decrease in FGF23 levels to 167.6 ± 5.57 pg/ml compared with the D₃-deficient group ($P < 0.001$). Upon induction of preeclampsia (Group IIIB), FGF23 levels increased to 386.5 ± 28.74 pg/ml compared with Group IIIA ($P < 0.01$), likely reflecting pronounced preeclampsia-associated renal dysfunction. In control rats, FGF23 levels remained unchanged during preeclampsia modeling (214.6 ± 14.99 pg/ml) compared with intact animals (208.2 ± 11.16 pg/ml).

The results of this study in an experimental rat pregnancy model confirm the pivotal role of vitamin D₃ in the hormonal regulation of calcium–phosphate metabolism under both physiological conditions and during the development of preeclampsia. The observed reduction in serum 25(OH)D₃ levels to nearly 16 ng/ml in vitamin D₃-deficient animals indicates the development of severe vitamin D₃ deficiency according to current classifications of vitamin D₃ status [24]. Previous studies have shown that low 25(OH)D₃ levels are associated with an increased risk of preeclampsia in pregnant women [25]. The 100% mortality observed in animals with experimentally induced preeclampsia on the background of severe vitamin D₃ deficiency highlights the compounded negative impact of these two factors (D₃ deficiency + PE), consistent with other experimental studies [26].

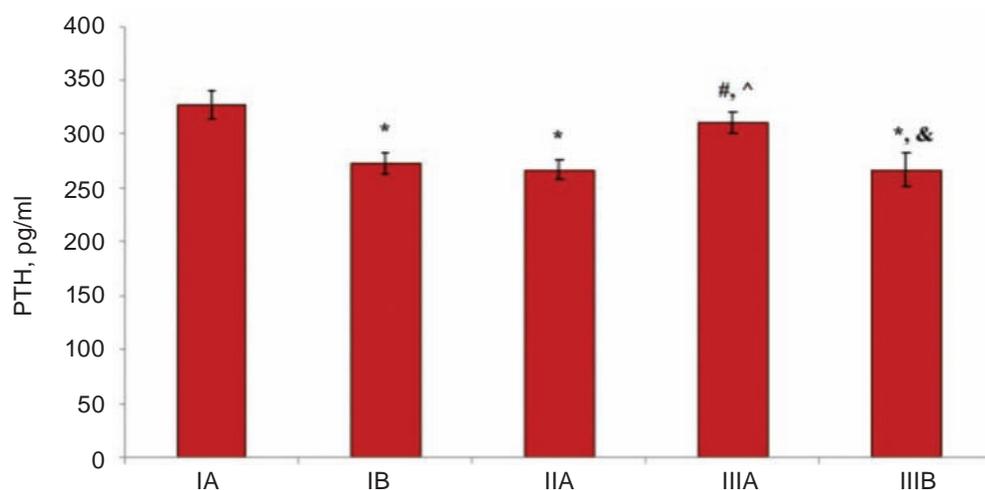


Fig. 4. Serum PTH levels in the studied groups ($M \pm m$): IA – control group without preeclampsia, IB – control group + preeclampsia, IIA – vitamin D₃-deficiency + without preeclampsia, IIIA – vitamin D₃-deficiency + vitamin D₃+ without preeclampsia, IIIB – vitamin D₃-deficiency + vitamin D₃ + preeclampsia. *Statistically significant difference vs. IA; #statistically significant difference vs. IIA; &statistically significant difference vs. IIIA; ^statistically significant difference vs. IB; $P < 0.05$

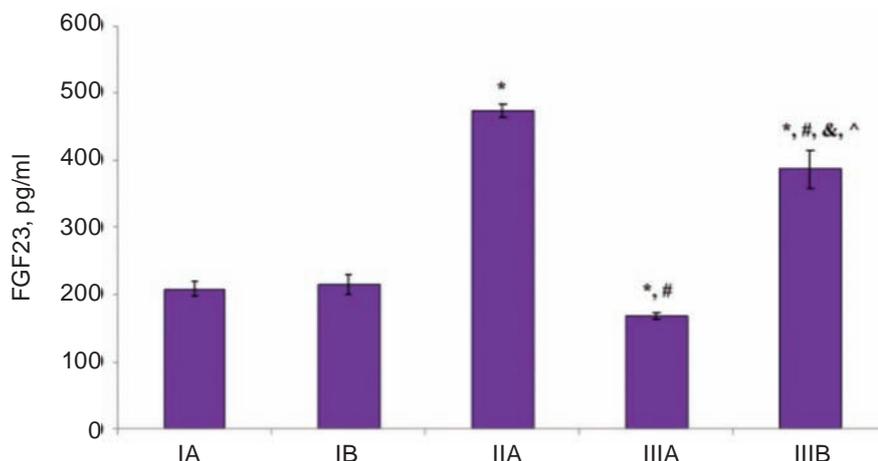


Fig. 5. Serum FGF23 levels in the studied groups ($M \pm m$): IA – control group without preeclampsia, IB – control group + preeclampsia, IIA – vitamin D₃-deficiency, without preeclampsia, IIIA – vitamin D₃-deficiency + vitamin D₃, without preeclampsia, IIIB – vitamin D₃-deficiency + vitamin D₃ + preeclampsia. *Statistically significant difference vs. IA; #statistically significant difference vs. IIA; &statistically significant difference vs. IIIA; ^statistically significant difference vs. IB; $P < 0.05$

Vitamin D₃ supplementation led to a significant increase in 25(OH)D₃ levels in Group IIIA, demonstrating the efficacy of the intervention and correlating with its important role in maintaining normal calcium/phosphate metabolism and placental function [27]. At the same time, the decrease in 25(OH)D₃ levels in Group IIIB (supplementation + PE) reflects a partial loss of corrective efficiency under more severe pathological conditions that is in agreement with experimental evidence of disrupted vitamin D₃ metabolism during preeclampsia, particularly due to inflammatory mechanisms and renal dysfunction [28-31].

Hemodynamic disturbances (elevated blood pressure) observed in the groups with vitamin D₃ deficiency and preeclampsia are consistent with studies indicating the role of vitamin D in regulating the renin-angiotensin-aldosterone system, endothelial function, and oxidative stress [30]. Vitamin D supplementation reduced blood pressure (Group IIIA), confirming its vasodilatory and anti-inflammatory effects [32]. However, the excessive increase in blood pressure observed during PE in Group IIIB emphasizes the complex pathophysiological and cellular-molecular mechanisms underlying preeclampsia pathogenesis, when the positive pharmacotherapeutic effects of vitamin D₃ may be limited by the severity of the pathological process [33].

Calcium metabolism exhibited classic features of hypocalcemia under D₃ deficiency and disrupted

mineral metabolism during PE, corroborating findings from numerous clinical and experimental studies [34, 35]. The increase in inorganic phosphate levels during PE indicates impaired renal excretion and activation of phosphaturic mechanisms, particularly mediated by hormonal influences of PTH and FGF23.

The classical (calcemic) role of vitamin D₃ is known to be primarily associated with the regulation of calcium-phosphate homeostasis along with two other key hormones produced by endocrine glands – parathyroid hormone (PTH) and calcitonin. Hypocalcemia and hypophosphatemia due to vitamin D₃ deficiency stimulate PTH secretion from the parathyroid glands, leading to secondary hyperparathyroidism. Increased synthesis and release of PTH enhance intestinal calcium absorption, bone mobilization through osteoclast-dependent resorption, and renal tubular reabsorption of calcium, while simultaneously increasing renal phosphate excretion [36-38]. Elevated PTH secretion in response to reduced serum calcium and phosphate additionally induces the expression of mitochondrial cytochrome P450 1 α -hydroxylase 25-hydroxyvitamin D (CYP27B1) in proximal renal tubular cells, initiating the synthesis of the biologically active metabolite calcitriol (1 α ,25(OH)₂D₃) – a high-affinity ligand that mediates both endocrine and auto/paracrine actions of this steroid hormone and serves as a potent regulator of phosphate-calcium homeostasis via the

VDR (vitamin D₃ receptor) transcription factor. The presence of secondary hyperparathyroidism may further indicate the development and severity of D₃ deficiency, as well as systemic disturbances in mineral metabolism.

It should be noted that, according to the literature, an increase in PTH levels during early pregnancy is generally a normal physiological adaptation to the additional calcium requirements of the fetus. Early pregnancy may mask symptoms of hyperparathyroidism caused by vitamin D₃ deficiency and hypocalcemia, which may explain the elevated PTH levels observed across all experimental groups in our study, with minor but statistically significant intergroup variations, either above or below those of the intact control group.

The role of FGF23 in the regulation of vitamin D₃ metabolism and mineral homeostasis during normal and pathological pregnancy remains an underexplored area. The alterations in FGF23 observed in our study closely correlate with the pathogenesis of phosphate metabolism disturbances. The significant increase in circulating FGF23 levels during vitamin D₃ deficiency may represent a protective compensatory response aimed at enhancing phosphaturia and reducing tissue calcification under conditions of intensified bone resorption [39].

The elevated FGF23 observed in preeclampsia (Group IIIB) reflects the severity of renal injury and exacerbated systemic inflammation, aligning with studies that identify FGF23 as a biomarker of renal dysfunction and cardiovascular risk [40-42]. Proteinuria, serving as an indicator of glomerular damage, was most pronounced in the PE groups, particularly when concomitant with vitamin D₃ deficiency. This finding is consistent with published literature and our previous data highlighting the role of vitamin D in maintaining renal function and preventing the progression of nephropathy during preeclampsia [43]. These observations underscore the interplay between vitamin D₃ status, phosphate-calcium homeostasis, and renal integrity under conditions of pathological pregnancy.

In summary, vitamin D₃ deficiency exacerbates the course of preeclampsia by intensifying hemodynamic disturbances, elevating FGF23 synthesis,

promoting phosphate-calcium imbalance, and impairing renal function. Correction of vitamin D₃ deficiency exerts beneficial effects on the investigated biochemical parameters and vascular function; however, its efficacy is limited in the presence of preeclampsia. These findings are of considerable relevance for future insight into the therapeutic potential of vitamin D₃ for the prevention and comprehensive management of preeclampsia.

Conclusions. This study demonstrated that vitamin D₃ deficiency in pregnant female rats leads to significant disturbances in calcium-phosphate homeostasis, increased serum levels of PTH and FGF23, and the associated activation of mechanisms contributing to hypertension. Vitamin D₃ supplementation partially restored vitamin D₃ status and mineral metabolism, reduced blood pressure, and mitigated renal injury in the absence of L-NAME-induced preeclampsia. However, the development of preeclampsia markedly worsened these parameters, exacerbating arterial hypertension and glomerular damage, highlighting the critical systemic role of vitamin D₃ deficiency in the pathogenesis of preeclampsia. The observed imbalance of regulatory factors governing calcium-phosphate metabolism under conditions of vitamin D₃ deficiency and experimental preeclampsia underscores the complex, multi-level mechanisms of hormonal and metabolic disturbances during pathological pregnancy.

Practical implications. The obtained results suggest the importance of early diagnosis and correction of vitamin D₃ deficiency to prevent the development of severe obstetric complications. Further studies should focus on elucidating the mechanisms of interaction between vitamin D₃ and multiple modulators of vascular-endothelial and renal function under conditions of preeclampsia complicated by vitamin D₃ deficiency and renal pathology.

Conflict of interest. Authors have completed the Unified Conflicts of Interest form at http://ukr-biochemjournal.org/wp-content/uploads/2018/12/coi_disclosure.pdf and declare no conflict of interest.

Funding. This work was funded by the National Research Foundation of Ukraine (project No. 2023.03/0222).

ФАКТОР РОСТУ ФІБРОБЛАСТІВ 23 ТА РІВНІ КАЛЬЦІЮ І ФОСФАТУ В СИРОВАТЦІ КРОВІ ЗА УМОВ ЕКСПЕРИМЕНТАЛЬНОЇ ПРЕЕКЛАМПСІЇ: ВПЛИВ СТАТУСУ ВІТАМІНУ D₃

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Преєклампсія (ПЕ) є однією з провідних причин материнської та перинатальної захворюваності, патогенез якої, зокрема, пов'язаний із порушенням мінерального гомеостазу. Останні дані свідчать, що фактор росту фібробластів 23 (FGF23) – ключовий регулятор фосфатного обміну та метаболізму вітаміну D₃ – може відігравати роль у розвитку акушерських ускладнень, однак його значення при ПЕ залишається недостатньо вивченим. Метою даного дослідження було оцінити рівень FGF23 у сироватці крові та його взаємозв'язок із вітаміном D₃ і кальцій-фосфатним балансом при розвитку преєклампсії в експериментальній тваринній моделі. Тридцять п'ять самок щурів лінії Wistar було розподілено на три групи: контрольну (стандартний раціон); щурів із дефіцитом вітаміну D₃; щурів із дефіцитом вітаміну D₃, яким додатково вводили холекальциферол. У кожній групі преєклампсію індукували шляхом введення N^ω-нітро-L-аргінін метилового ефіру. Концентрації 25(OH)D₃, FGF23, паратгормону (ПТГ), загального кальцію, неорганічного фосфату та активності лужної фосфатази (ЛФ) у сироватці крові визначали методом імуноферментного аналізу та біохімічних досліджень. Встановлено, що дефіцит вітаміну D₃ супроводжувався гіпокальціємією, гіпофосфатемією, підвищенням рівня FGF23 та зростанням активності ЛФ у сироватці крові. Додавання вітаміну D₃ призводило до підвищення рівня 25(OH)D₃, суттєвого зниження концентрації FGF23 та нормалізації мінеральних показників. Індукція преєклампсії зумовлювала виражені порушення кальцій-фосфатного обміну, розвиток артеріальної гіпертензії та 100%

летальність тварин із дефіцитом вітаміну D₃. За умов преєклампсії ефективність вітаміну D₃ була обмеженою. У групі ПЕ, незважаючи на застосування вітаміну D₃, рівень FGF23 у сироватці крові залишався значно підвищеним, що свідчить про порушення метаболізму вітаміну D₃. Отримані результати демонструють, що дефіцит вітаміну D₃ посилює тяжкість преєклампсії шляхом порушення мінерального гомеостазу та FGF23-залежної сигнальної регуляції.

Ключові слова: 25(OH)D₃, дефіцит вітаміну D₃, L-NAME, преєклампсія, кальцій-фосфатний обмін.

References

1. Rana S, Lemoine E, Granger JP, Karumanchi SA. Preeclampsia: Pathophysiology, Challenges, and Perspectives. *Circ Res.* 2019; 124(7): 1094-1112.
2. Opichka MA, Rappelt MW, Gutterman DD, Grobe JL, McIntosh JJ. Vascular Dysfunction in Preeclampsia. *Cells.* 2021; 10(11): 3055.
3. Moura TDB, Nunes FB, Crestani BDV, Araujo TFC, Hanauer EL, Corleta HVE, Branchini G. Preeclampsia and transport of ions and small molecules: A literature review. *Placenta.* 2024; 156: 77-91.
4. Javandoust Gharehbagh F, Soltani-Zangbar MS, Yousefzadeh Y. Immunological mechanisms in preeclampsia: A narrative review. *J Reprod Immunol.* 2024; 164: 104282.
5. Suresh S, Patel E, Mueller A, Morgan J, Lewandowski WL, Verlohren S, von Dadelszen P, Magee LA, Rana S. The additive role of angiogenic markers for women with confirmed preeclampsia. *Am J Obstet Gynecol.* 2023; 228(5): 573.e1-573.e11.
6. Abbasalizadeh S, Abam F, Mirghafourvand M, Abbasalizadeh F, Taghavi S, Hajizadeh K. Comparing levels of vitamin D, calcium and phosphorus in normotensive pregnant women and pregnant women with preeclampsia. *J Obstet Gynaecol.* 2020; 40(8): 1069-1073.
7. Bøllehuus Hansen L, Kaludjerovic J, Nielsen JE, Rehfeld A, Poulsen NN, Ide N, Skakkebaek NE, Frederiksen H, Juul A, Lanske B, Blomberg Jensen M. Influence of FGF23 and Klotho on male reproduction: Systemic vs direct effects. *FASEB J.* 2020; 34(9): 12436-12449.
8. Holick MF. The One-Hundred-Year Anniversary of the Discovery of the Sunshine Vitamin D₃:

- Historical, Personal Experience and Evidence-Based Perspectives. *Nutrients*. 2023; 15(3): 593.
9. Bikle DD. Vitamin D: Newer Concepts of Its Metabolism and Function at the Basic and Clinical Level. *J Endocr Soc*. 2020; 4(2): bvz038.
 10. Haussler MR, Livingston S, Sabir ZL, Haussler CA, Jurutka PW. Vitamin D Receptor Mediates a Myriad of Biological Actions Dependent on Its 1,25-Dihydroxyvitamin D Ligand: Distinct Regulatory Themes Revealed by Induction of Klotho and Fibroblast Growth Factor-23. *JBMR Plus*. 2020; 5(1): e10432.
 11. Carlberg C, Raczky M, Zawrotna N. Vitamin D: A master example of nutrigenomics. *Redox Biol*. 2023; 62: 102695.
 12. Rouhani P, Mokhtari E, Lotfi K, Saneei P. The association between circulating 25-hydroxyvitamin D levels and preeclampsia: a systematic review and dose-response meta-analysis of epidemiologic studies with GRADE assessment. *Nutr Rev*. 2023; 81(10): 1267-1289.
 13. Reddy M, Palmer K, Rolnik DL, Wallace EM, Mol BW, Da Silva Costa F. Role of placental, fetal and maternal cardiovascular markers in predicting adverse outcome in women with suspected or confirmed pre-eclampsia. *Ultrasound Obstet Gynecol*. 2022; 59(5): 596-605.
 14. Schröder-Heurich B, von Hardenberg S, Brodowski L, Kipke B, Meyer N, Borns K, von Kaisenberg CS, Brinkmann H, Claus P, von Versen-Höynck F. Vitamin D improves endothelial barrier integrity and counteracts inflammatory effects on endothelial progenitor cells. *FASEB J*. 2019; 33(8): 9142-9153.
 15. Grygorieva N, Tronko M, Kovalenko V, Komisarenko S, Tatarchuk T, Dedukh N, Veliky M, Strafun S, Komisarenko Y, Kalashnikov A, Orlenko V, Pankiv V, Shvets O, Gogunska I, Regeda S. Ukrainian Consensus on Diagnosis and Management of Vitamin D Deficiency in Adults. *Nutrients*. 2024; 16(2): 270.
 16. Mansur JL, Oliveri B, Giacoia E, Fusaro D, Costanzo PR. Vitamin D: Before, during and after Pregnancy: Effect on Neonates and Children. *Nutrients*. 2022; 14(9): 1900.
 17. You Z, Mei H, Zhang Y, Song D, Zhang Y, Liu C. The effect of vitamin D deficiency during pregnancy on adverse birth outcomes in neonates: a systematic review and meta-analysis. *Front Pediatr*. 2024; 12: 1399615.
 18. Shymanskyi IO, Lisakovska OO, Mazanova AO, Veliky MM. Vitamin D in transcriptional regulation of immune response and inflammation. *Advances in Medicine and Biology*. Ed. Leon V. Berhardt. NOVA science publisher; 2021; V. 183. P. 1-83.
 19. Kiely ME, Wagner CL, Roth DE. Vitamin D in pregnancy: Where we are and where we should go. *J Steroid Biochem Mol Biol*. 2020; 201: 105669.
 20. Stenhouse C, Halloran KM, Newton MG, Gaddy D, Suva LJ, Bazer FW. Novel mineral regulatory pathways in ovine pregnancy: I. phosphate, klotho signaling, and sodium-dependent phosphate transporters. *Biol Reprod*. 2021; 104(5): 1084-1096.
 21. Bakrania BA, George EM, Granger JP. Animal models of preeclampsia: investigating pathophysiology and therapeutic targets. *Am J Obstet Gynecol*. 2022; 226(2S): S973-S987.
 22. Stenhouse C, Halloran KM, Newton MG, Gaddy D, Suva LJ, Bazer FW. Novel mineral regulatory pathways in ovine pregnancy: II. Calcium-binding proteins, calcium transporters, and vitamin D signaling. *Biol Reprod*. 2021; 105(1): 232-243.
 23. Fahrleitner A, Dobnig H, Obernosterer A, Pilger E, Leb G, Weber K, Kudlacek S, Obermayer-Pietsch BM. Vitamin D deficiency and secondary hyperparathyroidism are common complications in patients with peripheral arterial disease. *J Gen Intern Med*. 2002; 17(9): 663-669.
 24. Kazemian E, Madreseh E, Azizi F, Ashrafiavand S, Gargari SS, Mansournia MA, Wagner CL, Amouzegar A. The association of parathyroid hormone with serum 25-hydroxyvitamin during pregnancy. *J Nutr Sci*. 2023; 12: e1.
 25. Wagner CL, Hollis BW. The extraordinary metabolism of vitamin D. *Elife*. 2022; 11: e77539.
 26. Durá-Travé T, Gallinas-Victoriano F. Pregnancy, Breastfeeding, and Vitamin D. *Int J Mol Sci*. 2023; 24(15): 11881.
 27. Vestergaard AL, Christensen M, Andreasen MF, Larsen A, Bor P. Vitamin D in pregnancy (GRAVITD) - a randomised controlled trial identifying associations and mechanisms linking maternal Vitamin D deficiency to placental dysfunction and adverse pregnancy outcomes - study protocol. *BMC Pregnancy Childbirth*. 2023; 23(1): 177.
 28. Vitoratos N, Lambrinouadaki I, Rizos D, Armeni E, Alexandrou A, Creatsas G. Maternal

- circulating osteoprotegerin and soluble RANKL in pre-eclamptic women. *Eur J Obstet Gynecol Reprod Biol.* 2011; 154(2): 141-145.
29. Benachi A, Baptiste A, Taieb J, Tsatsaris V, Guibourdenche J, Senat MV, Haidar H, Jani J, Guizani M, Jouannic JM, Haguet MC, Winer N, Masson D, Courbebaisse M, Elie C, Souberbielle JC. Relationship between vitamin D status in pregnancy and the risk for preeclampsia: A nested case-control study. *Clin Nutr.* 2020; 39(2): 440-446.
 30. Bi WG, Nuyt AM, Weiler H, Leduc L, Santamaria C, Wei SQ. Association Between Vitamin D Supplementation During Pregnancy and Offspring Growth, Morbidity, and Mortality: A Systematic Review and Meta-analysis. *JAMA Pediatr.* 2018; 172(7): 635-645.
 31. Lukyanova EM, Antipkin YuG, Omelchenko LI, Apukhovskaya LI. Vitamin D and its role in ensuring the health of children and pregnant women. Monograph. K. Iz-vo "Expert". 2005. 230 p.
 32. Poladych IV. Vitamin D in the genesis of preeclampsia: current understanding of the problem (literature review). *Bull Probl Biol Med.* 2024; 175(4): 113-122.
 33. Aouache R, Biquard L, Vaiman D, Miralles F. Oxidative Stress in Preeclampsia and Placental Diseases. *Int J Mol Sci.* 2018; 19(5): 1496.
 34. AlSubai A, Baqai MH, Agha H, Shankarlal N, Javaid SS, Jesrani EK, Golani S, Akram A, Qureshi F, Ahmed S, Saran S. Vitamin D and preeclampsia: A systematic review and meta-analysis. *SAGE Open Med.* 2023; 11: 20503121231212093.
 35. Shu W, Li H, Gong H, Zhang M, Niu X, Ma Y, Zhang X, Cai W, Yang G, Wei M, Yang N, Li Y. Evaluation of blood vessel injury, oxidative stress and circulating inflammatory factors in an L-NAME-induced preeclampsia-like rat model. *Exp Ther Med.* 2018; 16(2): 585-594.
 36. Nema J, Sundrani D, Joshi S. Prenatal vitamin D supplementation reduces blood pressure and improves placental angiogenesis in an animal model of preeclampsia. *Food Funct.* 2020; 11(12): 10413-10422.
 37. Sugulle M, Fiskå BS, Jacobsen DP, Fjeldstad HE, Staff AC. Placental Senescence and the Two-Stage Model of Preeclampsia. *Am J Reprod Immunol.* 2024; 92(1): e13904.
 38. Chau K, Welsh M, Makris A, Hennessy A. Progress in preeclampsia: the contribution of animal models. *J Hum Hypertens.* 2022; 36(8): 705-710.
 39. Riasniy VM, Apukhovska LI, Veliky NN, Shymanskyi IO, Labudzynskyi DO, Komisarenko SV. Immunomodulatory effects of vitamin D₃ and bisphosphonates in nutritional osteoporosis in rats. *Ukr Biokhim Zhurn.* 2012; 84(2): 73-80. (In Ukrainian).
 40. Ramdin S, Baijnath S, Naicker T, Govender N. The Clinical Value of Rodent Models in Understanding Preeclampsia Development and Progression. *Curr Hypertens Rep.* 2023; 25(6): 77-89.
 41. Xie H, Bastepe I, Zhou W, Ay B, Ceraj Z, Portales-Castillo IA, Liu ES, Burnett-Bowie SM, Jüppner H, Rhee EP, Bastepe M, Simic P. 1,25-Dihydroxyvitamin D₃ regulates furin-mediated FGF23 cleavage. *JCI Insight.* 2023; 8(17): e168957.
 42. Martínez-Heredia L, Canelo-Moreno JM, García-Fontana B, Muñoz-Torres M. Non-Classical Effects of FGF23: Molecular and Clinical Features. *Int J Mol Sci.* 2024; 25(9): 4875.
 43. Poladych IV, Shymanskyi IO, Veliky MM, Govsieiev DO. Experimental preeclampsia development depends on vitamin D₃ status in female wistar rats. *Ukr Biochem J.* 2025; 97(4): 34-42.